## Pancreas

# Digestive Diseases

Dig Dis 2016;34:525–531 DOI: 10.1159/000445233

# Development of Pancreatic Cancer: Targets for Early Detection and Treatment

Markus M. Lerch Julia Mayerle Ujjwal Mahajan Matthias Sendler F. Ulrich Weiss Ali Aghdassi Patryk Moskwa Peter Simon

Department of Medicine A, University Medicine Greifswald, Greifswald, Germany

#### **Key Words**

Biomarkers · Extracellular matrix · Pancreatic cancer · Pancreatitis · Precision medicine

#### Abstract

Background: Pancreatic ductal adenocarcinoma (PDAC) is the 4th leading cause of cancer death worldwide and compared to other malignancies its share in cancer mortality is expected to rise further. This is due to a lack of sensitive diagnostic tools that would permit earlier detection in a potentially curable stage and the very slow progress in finding effective drug treatments for pancreatic cancer. Key Messages: Aside from genetic predispositions and environmental agents, chronic pancreatitis is by far the greatest risk factor for PDAC. It also shares several etiological factors with pancreatic cancer and represents its most challenging differential diagnosis. Biomarkers that can distinguish between chronic pancreatitis and PDAC may therefore be suitable for the latter's early detection. Moreover, targeting the natural history of chronic pancreatitis would be one approach to prevent PDAC. Targeting tumor-cell signaling directly by interfering with receptor tyrosine kinases has shown some efficacy, although the results in clinical trials were less encouraging than for other cancers. Other compounds developed have targeted the formation of extracellular matrix around the tumor, the proteolytic activity in the tumor environment, histone deacetylases, hedgehog signaling and heat shock

# KARGER

© 2016 S. Karger AG, Basel 0257-2753/16/0345-0525\$39.50/0

E-Mail karger@karger.com www.karger.com/ddi proteins, but none has yet found its way into routine patient care. Attempts to individualize treatment according to the tumor's somatic mutation profile are novel but so far impractical. **Conclusions:** Progress in the treatment of pancreatic cancer has been exceedingly slow and mostly dependent on improved pharmaceutical preparations or combinations of established chemotherapeutic agents. The promise of major breakthroughs implied in targeting tumor signal transduction events has so far not materialized. © 2016 S. Karger AG, Basel

#### Introduction

Pancreatic cancer is currently the 4th leading cause of cancer death and projected to become the 3rd leading cause of cancer-related death by 2030 due to delayed diagnosis and slow progress in treatment development [1]. The dismal prognosis of pancreatic cancer is caused by a variety of factors: (a) due to its location in the retroperitoneum, pancreatic cancer causes symptoms only when it has already grown to an advanced stage and is no longer locally resectable [2]. (b) Pancreatic cancer has a tendency to disseminate not only into the bloodstream and into the lymphatic tissue but also along nerve fibers, leading to an unusually high recurrence rate even after successful R0 resection [3]. For both these issues, a current consensus predicts that methods allowing earlier detection of pan-

Markus M. Lerch, MD, FRCP Department of Medicine A University Medicine Greifswald Ferdinand-Sauerbruch-Strasse, DE–17475 Greifswald (Germany) E-Mail lerch@uni-greifswald.de creatic cancer would be of benefit and improve survival [4]. (c) Pancreatic cancer is highly resistant to chemotherapy, radiation therapy and even targeted therapy [5].

#### **Early Detection of Pancreatic Cancer**

In the context of early detection methods and outside of imaging technologies, only very few biomarkers have been identified so far that could either distinguish pancreatic cancer from other disorders of the pancreas or detect it earlier than with currently available methods. The best established blood test for this purpose is carbohydrate antigen 19-9 (CA19-9), a Lewis antigen of the MUC1 protein-class. Unfortunately, CA19-9 can also be elevated in patients with nonmalignant diseases including liver cirrhosis, chronic pancreatitis and cholangitis as well as other gastrointestinal cancers [6]. CA19-9 has been reported to discriminate between pancreatic cancer patients and healthy controls with a sensitivity and specificity of slightly over 80% [7] and between pancreatic ductal adenocarcinoma (PDAC) and benign pancreatic disease with a sensitivity of 78% and a specificity of 83% [8]. However, CA19-9 is not expressed in Lewis blood type negative patients and this limits the optimum of any test relying on CA19-9 at a sensitivity of 92% under the best of circumstances. In up to one third of patients, the distinction between chronic pancreatitis and PDAC is inaccurate and the negative predictive value of diagnostic assays is often no better than 50-60% [1]. This has prompted a search for diagnostic biomarkers in blood and other body fluids (including saliva) for 2 purposes: to distinguish between benign pancreatic disease and pancreatic cancer and to detect PDAC earlier than currently possible with established imaging techniques. One very promising step in this quest is the detection of exosomes or microRNA patterns with a reasonable degree of specificity for PDAC [9]. Our group has taken a different approach and searched for metabolic biomarkers using a metabolomics approach including lipidomics. In more than 900 patients and appropriate controls, we were able to identify a distinct biomarker signature that can distinguish pancreatic cancer from chronic pancreatitis with greater sensitivity and specificity than CA19-9 and would improve the accuracy of the detection in 30% of patients [10]. Biomarker signatures (panels of multiple markers), rather than single individual parameters appear presently the most promising approach to early pancreatic cancer detection and thus the diagnosis in a potentially curable stage. Whether they can be employed in a populationbased screening approach rather than being used only to make the distinction between clinically manifest benign and malignant pancreatic disorders has not been established and requires further studies.

#### **Treatments Resistance of Pancreatic Cancer**

The issue of therapy resistance is even harder to address. Some progress has been made over recent decades in identifying chemotherapy regimens that increase the overall survival of patients with pancreatic cancer. The most notable success was to establish that adjuvant chemotherapy in pancreatic and ampullary cancer can double the survival of patients after successful resection of the tumor [11, 12]. However, the fact that this merely represents an improvement from 10% to approximately 20% overall survival after 5 years only highlights the extensive treatment resistance and almost limitless recurrence potential of this tumor. For patients in whom surgery is not an option because the tumor is locally advanced, usually with artery encasement beyond a resectable stage, or because of metastasis formation, palliative chemotherapy is currently the only therapeutic option. Improvements in overall survival have been achieved in recent years, albeit the extent is limited. One regimen that was found to confer a survival benefit was the combination of several long established cytotoxic compounds such as 5FU, irinotecan and oxaliplatin [13]. Another used a new albumin-encapsulated preparation of paclitaxel in combination with gemcitabine and found it superior to the long-standing standard regimen of gemcitabine alone [14]. A similar approach to improving the tumor penetrance of established chemotherapeutic agents was taken with irinotecan, a liposomal preparation of which has resulted in encouraging initial studies [15] and will probably be approved for pancreatic cancer within the next months. These regimens have improved the median overall survival from 5 months to no more than 11 months and thus represent a much lesser treatment advance than achieved for patients with colorectal cancer or other solid tumors. A summary of currently available treatment options can be found in table 1.

#### **Potential and Established Treatment Targets**

A variety of structures have been targeted in pancreatic cancer such as the EGF receptor, the VEGF receptor, fibroblast activation protein  $\alpha 5\beta 1$ -Integin, and others **Table 1.** Life expectancy in patients with pancreatic cancer

5-year overall survival of all patients, %	0.4
5-year overall survival after resection and adjuvant chemotherapy, %	20
Median survival, months	
All patients, best supportive care	5
Chemotherapy with gemcitabine	6
Chemotherapy with gemcitabine plus capecitabine	7
Chemotherapy with gemcitabine plus nab-paclitaxel	9
Chemotherapy with gemcitabine plus erlotinib (in case of rash)	10
Chemotherapy with FOLFIRINOX	11
Resection without adjuvant chemotherapy	16
Resection with adjuvant chemotherapy (gemcitabine or 5 fluoruracil)	23
Current average gain in life expectancy by surgery, months	12–17
Gudjonsson B: Cancer 1987;60:2284–2303 [52] Neoptolemos JP, et al: N Engl J Med 2004;350:1200–1210 [53] Moore MJ et al: J Clin Oncol 2007;25:1960–1966 [16] Conroy T, et al: N Engl J Med 2011;364:1817–1825 [13]	Neoptolemos JP, et al: JAMA 2010;304:1073–1081 [11] Burris HA 3rd, et al: J Clin Oncol 1997;15:2403–2413 [54] Cunningham D, et al: J Clin Oncol 2009;27:5513–5518 [55] Goldstein D, et al: J Natl Cancer Inst 2015;pii:dju413 [14]

without resulting in a significant clinical benefit. Neither overexpression of TNFa nor the use of broad spectrum receptor tyrosine kinase inhibitors has met the high expectations of patients and physicians. The only tyrosine kinase inhibitor found to confer a survival advantage so far was erlotinib, a small molecule originally designed to interfere with EGF receptor signaling but probably also effective against more than 20 other RTKs [16]. Not every patient benefits from treatment with erlotinib but only those who develop a prominent skin rash. The skin rash therefore serves as a biomarker sign and can be used to determine whether or not a continuation of the treatment beyond a few weeks is of any benefit to the patient. Other RTKs [17] are currently under investigation as potential treatment targets and it is much hoped that novel compounds directed against them can match or surpass the results achieved with erlotinib.

Since more than 90% of pancreatic cancer specimens carry somatic mutations in the proto-oncogene *KRAS* this has turned into an attractive target. The disadvantage of the RAS pathway is that it is vital for cellular survival in all tissues and not only cancer cells. This poses a number of difficulties for developing tumor-specific anti-RAS therapies. Other targets that are currently under investigation include polo-like kinase and heat shock protein 70, the latter of which appears to be involved in tumor cell resistance to apoptosis [18]. The dissemination of tumor cells into neighboring organs depends on the function of intact cell–cell adhesions [2], which impair metastasis formation. Histone deacetylases HDAC1 and HDAC2

Development of Pancreatic Cancer: Targets for Early Detection and Treatment are potent regulators of cell-contact protein formation and have therefore (including their inhibitors) become a much investigated target for pancreatic cancer treatment modalities [19]. The jury is still out as to whether or not they are of any benefit to patients. Once the appropriate target has been identified, there remains the challenge of how to deliver the compound to the tumor cells in a cancer that produces abundant extracellular matrix, an often severe impediment to drug delivery. To address this challenge a number of techniques including microspheres and the abovementioned albumin- or liposome encapsulation of compounds have been invented. Other strategies target the tumor stoma directly.

#### **Targeting Extracellular Matrix Deposition**

One of the well-researched explanations why pancreatic cancer is so resistant to chemotherapy is the fact that it produces extensive extracellular matrix [20] encapsulating the tumor, a phenomenon it shares with chronic pancreatitis [21], and that any systemically administered medication can simply not reach the tumor across this barrier [22]. This assumption has prompted the development of several strategies intended to overcome the matrix barrier and to degrade or digest its components in pancreatic cancer [23]. Recent experimental evidence suggests that this may not necessarily be beneficial and could even render the exposed tumor cells more aggressive [24, 25]. Inhibition of the hedgehog pathway, regarded as crucial for pancreatic cancer desmoplasia and matrix deposition, also resulted in disappointing outcomes [26]. The longstanding discussion of whether the extracellular matrix protects the patient from a dissemination of his tumor or whether it protects the tumor from the penetration of chemotherapeutic agents or tumor-lytic inflammatory cells appears not to be settled for the moment.

### **Precision Therapy**

Pancreatic cancer is known to carry a multitude of somatic mutations affecting a finite number of signal transduction pathways. A strategy has therefore been developed; it is broadly based on the concept of individualized medicine, more recently termed precision medicine. The approach involves taking a tumor biopsy, analyzing the tumor genome and characterizing the signaling pathways that have undergone pathological alterations. Based on these findings, an individually confectioned cocktail of antiproliferative agents or inhibitors of signal transduction pathways shall then be administered. Investigators have so far screened for established molecular targets such as HER2 amplification, KRAS wild-type, and mutations in DNA damage repair pathways (BRCA1, BRCA2, PALB2, ATM). The first pilot results are not encouraging but the greatest impediments are mostly technical. They include the need to obtain a sufficient number of tumor cells on biopsy or the delay of 3 weeks until final results are translated into a therapy, which resulted in an unacceptable dropout rate in one study [27]. However, these difficulties will most likely be overcome by technical improvements and only then can individualization of therapy according to the genomic tumor profile be assessed with a sufficient degree of robustness.

# Pancreatic Cancer and Chronic Pancreatitis

Chronic pancreatitis is not only the single most significant risk factor for the development of pancreatic cancer but also an important differential diagnosis [28]. Particularly patients suffering from the hereditary variety of chronic pancreatitis that is associated with mutations in the cationic trypsinogen (*PRSS1*) gene have a 40–70% lifetime risk of developing pancreatic cancer [29]. They further double their cancer risk if additional environmental factors such as cigarette smoking contribute to this condition [30]. It seems presently unlikely that acute pancreatitis [31] can contribute to the pancreatic cancer risk, but for autoimmune pancreatitis, a possible association has not been ruled out [32, 33].

Chronic pancreatitis and pancreatic cancer share a number of underlying mechanisms and risk factors. Common to all forms of pancreatitis is a prominent role of trypsin, a digestive protease that is prematurely activated in the early disease phase [34, 35] and mutations in one of the isoforms of which (PRSS1) confer the greatest risk of developing hereditary pancreatitis [28, 29], the disease variety burdened with the greatest risk of developing pancreatic cancer [29, 30]. Interestingly, trypsin is also immunogenic and has been implicated in the pathogenesis of autoimmune pancreatitis [36]. The role of trypsin in pancreatic cancer is less clear, but several studies have implicated this digestive protease as either a biomarker [37] or a pathogenetic factor [38] for adenocarcinoma of the pancreas. The same connection applies to another group of proteases, lysosomal cathepsins, secreted by the exocrine pancreas in significant quantities [39, 40], play a prominent role in the activation of trypsin during pancreatitis [41, 42], but have also been identified in pancreatic cancer tissue, where their presence appears to be a biomarker for a poor prognosis [43]. Their pathogenetic role in cancer is assumed to require an involvement in the interaction between cancer cells and extracellular matrix.

Another common mechanism between pancreatitis and cancer deals with the functional impairment of cell– cell contacts [44], which not only permits the translocation of inflammatory cells into the pancreas [45, 46], but at the same time allows for the dissemination of malignant cells from the tumor to peripheral organs [2]. Among the cell contact, protein families that have been shown to be involved in tumor development are claudins [47], mutations in which have recently been reported to also represent a risk factor for chronic pancreatitis [48].

The last factor on this incomplete list worth mentioning, and presently the most puzzling, is the ABO blood type. Blood types are a surrogate for the degree to which certain cellular proteins, and not only those on red blood cells, undergo surface glycosylation. It has been shown that the blood type B not only increases the lifetime risk of developing pancreatic cancer [49], but also doubles the risk of developing chronic pancreatitis [50]. Whether or not the underlying mechanisms indicated by blood type B only signals an unspecific cellular stress (ER-stress) response as common denominator for pancreatitis and cancer or rather points to a distinct set of proteins that need to undergo specific glycosylation events for the disease risk to materialize is presently unknown. Studies that attempt to solve this question are ongoing.

At present it appears plausible that any target that is common to pancreatitis and pancreatic cancer has the greatest potential for leading to a preventive strategy. As shown for other inflammatory disorders such as ulcerative colitis and hepatitis, it is likely that approaches with a beneficial effect on the natural disease course of pancreatitis will have an impact on the development of pancreatic cancer. Currently there is none, and therapy for chronic pancreatitis remains strictly symptomatic or complication oriented. This needs to change before greater progress in the management of patients with pancreatitis and in the prevention of pancreatic cancer can be expected [51]. Pancreatic cancer surgery is, in all but a tiny minority of patients, as much a palliative approach as conventional chemotherapy. It is at present just the better palliation and prolongs the patients live significantly longer. For medical therapy of pancreatic cancer to be more successful, two issues need to be resolved: (a) how to target tumor cells or tumor stem cells more specifically, effectively and sustainably and (b) how to deliver this therapy to a tumor across a highly impenetrable extracellular matrix barrier. A large-scale concerted ef-

#### References

- 1 Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM: Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. Cancer Res 2014;74:2913–2921.
- 2 Mayerle J, Friess H, Büchler MW, Schnekenburger J, Weiss FU, Zimmer KP, Domschke W, Lerch MM: Up-regulation, nuclear import, and tumor growth stimulation of the adhesion protein p120 in pancreatic cancer. Gastroenterology 2003;124:949–960.
- 3 Seufferlein T, Porzner M, Becker T, Budach V, Ceyhan G, Esposito I, Fietkau R, Follmann M, Friess H, Galle P, Geissler M, Glanemann M, Gress T, Heinemann V, Hohenberger W, Hopt U, Izbicki J, Klar E, Kleeff J, Kopp I, Kullmann F, Langer T, Langrehr J, Lerch M, Löhr M, Lüttges J, Lutz M, Mayerle J, Michl P, Möller P, Molls M, Münter M, Nothacker M, Oettle H, Post S, Reinacher-Schick A, Röcken C, Roeb E, Saeger H, Schmid R, Schmiegel W, Schoenberg M, Siveke J, Stuschke M, Tannapfel A, Uhl W, Unverzagt S, van Oorschot B, Vashist Y, Werner J, Yekebas E; Guidelines Programme Oncology AWMF; German Cancer Society eV; German Cancer Aid: [S3guideline exocrine pancreatic cancer]. Z Gastroenterol 2013;51:1395-1440.
- 4 Chari ST, Kelly K, Hollingsworth MA, Thayer SP, Ahlquist DA, Andersen DK, Batra SK, Brentnall TA, Canto M, Cleeter DF, Firpo MA, Gambhir SS, Go VL, Hines OJ, Kenner BJ, Klimstra DS, Lerch MM, Levy MJ, Maitra A, Mulvihill SJ, Petersen GM, Rhim AD, Simeone DM, Srivastava S, Tanaka M, Vinik AI, Wong D: Early detection of sporadic pancreatic cancer: summative review. Pancreas 2015;44:693–712.
- 5 Nambaru PK, Hübner T, Köck K, Mews S, Grube M, Payen L, Guitton J, Sendler M, Jedlitschky G, Rimmbach C, Rosskopf D, Kowalczyk DW, Kroemer HK, Weiss FU, Mayerle J, Lerch MM, Ritter CA: Drug efflux transporter multidrug resistance-associated protein 5 affects sensitivity of pancreatic cancer cell lines to the nucleoside anticancer drug 5-fluorouracil. Drug Metab Dispos 2011;39:132–139.
- 6 Duffy MJ, Sturgeon C, Lamerz R, Haglund C, Holubec VL, Klapdor R, Nicolini A, Topolcan O, Heinemann V: Tumor markers in pancreatic cancer: a European group on tumor markers (EGTM) status report. Ann Oncol 2010;21:441–447.
- 7 Gui JC, Yan WL, Liu XD: CA19–9 and CA242 as tumor markers for the diagnosis of pancreatic cancer: a meta-analysis. Clin Exp Med 2014;14:225–233.

fort by the research community and its funding agencies is required to make this deadly disease not only a nonsurgical disorder but also a curable condition.

#### Acknowledgments

Dedicated to Professor Wolfgang Gerok, Freiburg, on the occasion of his 90th birthday.

The authors' original work was supported by grants from Deutsche Krebshilfe/Dr. Mildred-Scheel-Stiftung (109102), the Deutsche Forschungsgemeinschaft (DFG GRK840-D2/E3/E4, MA 4115/1-2/3, AG 203/2-1, MO 2924/1-1), the Federal Ministry of Education and Research (BMBF GANI-MED 03IS2061A and BMBF 0314107, 01ZZ9603, 01ZZ0103, 01ZZ0403, 03ZIK012) the European Union (EU-FP-7: EPC-TM and EU-FP7-REG-POT-2010-1) and EFRE-State Ministry of Economics MV (V-630-S-150-2012/132/133).

#### **Disclosure Statement**

M.M.L. has served on advisory boards for ISIS, Roche, Nordmark, Abbvie, Abbott, Centogene and Astra-Zeneca and received grant support from Celgene, Sanofi-Aventis, Novartis and Astra-Zeneca. M.M.L. and J.M. hold patents on metabolomic biomarker panels with Metanomics Health.

- 8 Poruk KE, Gay DZ, Brown K, Mulvihill JD, Boucher KM, Scaife CL, Firpo MA, Mulvihill SJ: The clinical utility of CA 19–9 in pancreatic adenocarcinoma: diagnostic and prognostic updates. Curr Mol Med 2013;13:340–351.
- 9 Schultz NA, Dehlendorff C, Jensen BV, Bjerregaard JK, Nielsen KR, Bojesen SE, Calatayud D, Nielsen SE, Yilmaz M, Holländer NH, Andersen KK, Johansen JS: MicroRNA biomarkers in whole blood for detection of pancreatic cancer. JAMA 2014;311:392–404.
- 10 Mayerle J, Kalthoff H, Reszka R, Kamlage B, Peter E, Schniewind B, Gonzalez-Maldonado S, Liebenberg V, Pilarsky C, Schatz P, Scheiber JA, Weiss FU, Grützmann R, Lerch MM: Metabolic biomarkers for the diagnosis of pancreatic ductal adenocarcinoma. Pancreatology 2014;14:S19.
- 1 Neoptolemos JP, Stocken DD, Bassi C, Ghaneh P, Cunningham D, Goldstein D, Padbury R, Moore MJ, Gallinger S, Mariette C, Wente MN, Izbicki JR, Friess H, Lerch MM, Dervenis C, Oláh A, Butturini G, Doi R, Lind PA, Smith D, Valle JW, Palmer DH, Buckels JA, Thompson J, McKay CJ, Rawcliffe CL, Büchler MW; European Study Group for Pancreatic Cancer: Adjuvant chemotherapy with fluorouracil plus folinic acid vs gemcitabine following pancreatic cancer resection: a randomized controlled trial. JAMA 2010;304:1073–1081.

- 12 Neoptolemos JP, Moore MJ, Cox TF, Valle JW, Palmer DH, McDonald AC, Carter R, Tebbutt NC, Dervenis C, Smith D, Glimelius B, Charnley RM, Lacaine F, Scarfe AG, Middleton MR, Anthoney A, Ghaneh P, Halloran CM, Lerch MM, Oláh A, Rawcliffe CL, Verbeke CS, Campbell F, Büchler MW; European Study Group for Pancreatic Cancer: Effect of adjuvant chemotherapy with fluorouracil plus folinic acid or gemcitabine vs observation on survival in patients with resected periampullary adenocarcinoma: the ES-PAC-3 periampullary cancer randomized trial. JAMA 2012;308:147–156.
- 13 Conroy T, Desseigne F, Ychou M, Bouché O, Guimbaud R, Bécouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardière C, Bennouna J, Bachet JB, Khemissa-Akouz F, Péré-Vergé D, Delbaldo C, Assenat E, Chauffert B, Michel P, Montoto-Grillot C, Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup: FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. N Engl J Med 2011;364:1817–1825.
- 14 Goldstein D, El-Maraghi RH, Hammel P, Heinemann V, Kunzmann V, Sastre J, Scheithauer W, Siena S, Tabernero J, Teixeira L, Tortora G, Van Laethem JL, Young R, Penenberg DN, Lu B, Romano A, Von Hoff DD: Nab-paclitaxel plus gemcitabine for metastatic pancreatic cancer: long-term survival from a phase III trial. J Natl Cancer Inst 2015;pii:dju413.
- 15 Ko AH, Tempero MA, Shan YS, Su WC, Lin YL, Dito E, Ong A, Wang YW, Yeh CG, Chen LT: A multinational phase 2 study of nanoliposomal irinotecan sucrosofate (PEP02, MM-398) for patients with gemcitabine-refractory metastatic pancreatic cancer. Br J Cancer 2013;109:920–925.
- 16 Moore MJ, Goldstein D, Hamm J, Figer A, Hecht JR, Gallinger S, Au HJ, Murawa P, Walde D, Wolff RA, Campos D, Lim R, Ding K, Clark G, Voskoglou-Nomikos T, Ptasynski M, Parulekar W; National Cancer Institute of Canada Clinical Trials Group: Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. J Clin Oncol 2007;25:1960–1966.
- 17 Ciossek T, Lerch MM, Ullrich A: Cloning, characterization, and differential expression of MDK2 and MDK5, two novel receptor tyrosine kinases of the eck/eph family. Oncogene 1995;11:2085–2095.
- 18 Aghdassi A, Phillips P, Dudeja V, Dhaulakhandi D, Sharif R, Dawra R, Lerch MM, Saluja A: Heat shock protein 70 increases tumorigenicity and inhibits apoptosis in pancreatic adenocarcinoma. Cancer Res 2007;67:616–625.
- 19 Aghdassi A, Sendler M, Guenther A, Mayerle J, Behn CO, Heidecke CD, Friess H, Büchler M, Evert M, Lerch MM, Weiss FU: Recruitment of histone deacetylases HDAC1 and HDAC2 by the transcriptional repressor ZEB1 downregulates E-cadherin expres-

sion in pancreatic cancer. Gut 2012;61:439-448.

- 20 Gress TM, Müller-Pillasch F, Lerch MM, Friess H, Büchler M, Adler G: Expression and in-situ localization of genes coding for extracellular matrix proteins and extracellular matrix degrading proteases in pancreatic cancer. Int J Cancer 1995;62:407–413.
- 21 Gress TM, Müller-Pillasch F, Lerch MM, Friess H, Büchler M, Beger HG, Adler G: Balance of expression of genes coding for extracellular matrix proteins and extracellular matrix degrading proteases in chronic pancreatitis. Z Gastroenterol 1994;32:221–225.
- 22 Neesse A, Krug S, Gress TM, Tuveson DA, Michl P: Emerging concepts in pancreatic cancer medicine: targeting the tumor stroma. Onco Targets Ther 2013;7:33–43.
- 23 Provenzano PP, Cuevas C, Chang AE, Goel VK, Von Hoff DD, Hingorani SR: Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma. Cancer Cell 2012;21:418–429.
- 24 Rhim AD, Oberstein PE, Thomas DH, Mirek ET, Palermo CF, Sastra SA, Dekleva EN, Saunders T, Becerra CP, Tattersall IW, Westphalen CB, Kitajewski J, Fernandez-Barrena MG, Fernandez-Zapico ME, Iacobuzio-Donahue C, Olive KP, Stanger BZ: Stromal elements act to restrain, rather than support, pancreatic ductal adenocarcinoma. Cancer Cell 2014;25:735–747.
- 25 Özdemir BC, Pentcheva-Hoang T, Carstens JL, Zheng X, Wu CC, Simpson TR, Laklai H, Sugimoto H, Kahlert C, Novitskiy SV, De Jesus-Acosta A, Sharma P, Heidari P, Mahmood U, Chin L, Moses HL, Weaver VM, Maitra A, Allison JP, LeBleu VS, Kalluri R: Depletion of carcinoma-associated fibroblasts and fibrosis induces immunosuppression and accelerates pancreas cancer with reduced survival. Cancer Cell 2014;25:719–734.
- 26 Ko AH1, LoConte N, Tempero MA, Walker EJ, Kate Kelley R, Lewis S, Chang WC, Kantoff E, Vannier MW, Catenacci DV, Venook AP, Kindler HL: A phase I study of FOLFIRINOX Plus IPI-926, a Hedgehog Pathway Inhibitor, for Advanced pancreatic adenocarcinoma. Pancreas 2016;45:370–375.
- 27 Chantrill LA, Nagrial AM, Watson C, Johns AL, Martyn-Smith M, Simpson S, Mead S, Jones MD, Samra JS, Gill AJ, Watson N, Chin VT, Humphris JL, Chou A, Brown B, Morey A, Pajic M, Grimmond SM, Chang DK, Thomas D, Sebastian L, Sjoquist K, Yip S, Pavlakis N, Asghari R, Harvey S, Grimison P, Simes J, Biankin AV; Australian Pancreatic Cancer Genome Initiative (APGI); Individualized Molecular Pancreatic Cancer Therapy (IMPaCT) Trial Management Committee of the Australasian Gastrointestinal Trials Group (AGITG): Precision medicine for advanced pancreas cancer: the individualized molecular pancreatic cancer therapy (IMPaCT) trial. Clin Cancer Res 2015;21: 2029-2037.

- 28 Ellis I, Lerch MM, Whitcomb DC; Consensus Committees of the European Registry of Hereditary Pancreatic Diseases, Midwest Multi-Center Pancreatic Study Group, International Association of Pancreatology: Genetic testing for hereditary pancreatitis: guidelines for indications, counselling, consent and privacy issues. Pancreatology 2001; 1:405–415.
- 29 Keim V, Bauer N, Teich N, Simon P, Lerch MM, Mössner J: Clinical characterization of patients with hereditary pancreatitis and mutations in the cationic trypsinogen gene. Am J Med 2001;111:622–626.
- 30 Lowenfels AB, Maisonneuve P, Whitcomb DC, Lerch MM, DiMagno EP: Cigarette smoking as a risk factor for pancreatic cancer in patients with hereditary pancreatitis. JAMA 2001;286:169–170.
- 31 Hernández CA, Lerch MM: Sphincter stenosis and gallstone migration through the biliary tract. Lancet 1993;341:1371–1373.
- 32 Hart PA, Kamisawa T, Brugge WR, Chung JB, Culver EL, Czakó L, Frulloni L, Go VL, Gress TM, Kim MH, Kawa S, Lee KT, Lerch MM, Liao WC, Löhr M, Okazaki K, Ryu JK, Schleinitz N, Shimizu K, Shimosegawa T, Soetikno R, Webster G, Yadav D, Zen Y, Chari ST: Long-term outcomes of autoimmune pancreatitis: a multicentre, international analysis. Gut 2013;62:1771–1776.
- 33 Kamisawa T, Chari ST, Giday SA, Kim MH, Chung JB, Lee KT, Werner J, Bergmann F, Lerch MM, Mayerle J, Pickartz T, Lohr M, Schneider A, Frulloni L, Webster GJ, Reddy DN, Liao WC, Wang HP, Okazaki K, Shimosegawa T, Kloeppel G, Go VL: Clinical profile of autoimmune pancreatitis and its histological subtypes: an international multicenter survey. Pancreas 2011;40:809–814.
- 34 Krüger B, Lerch MM, Tessenow W: Direct detection of premature protease activation in living pancreatic acinar cells. Lab Invest 1998; 78:763–764.
- 35 Halangk W, Krüger B, Ruthenbürger M, Stürzebecher J, Albrecht E, Lippert H, Lerch MM: Trypsin activity is not involved in premature, intrapancreatic trypsinogen activation. Am J Physiol Gastrointest Liver Physiol 2002; 282:G367–G374.
- 36 Löhr JM, Faissner R, Koczan D, Bewerunge P, Bassi C, Brors B, Eils R, Frulloni L, Funk A, Halangk W, Jesenofsky R, Kaderali L, Kleeff J, Krüger B, Lerch MM, Lösel R, Magnani M, Neumaier M, Nittka S, Sahin-Tóth M, Sänger J, Serafini S, Schnölzer M, Thierse HJ, Wandschneider S, Zamboni G, Klöppel G: Autoantibodies against the exocrine pancreas in autoimmune pancreatitis: gene and protein expression profiling and immunoassays identify pancreatic enzymes as a major target of the inflammatory process. Am J Gastroenterol 2010;105:2060–2071.
- 37 Lake-Bakaar G, McKavanagh S, Summerfield JA: Urinary immunoreactive trypsin excretion: a non-invasive screening test for pancreatic cancer. Lancet 1979;2:878–880.

- 38 Johansen D, Manjer J, Regner S, Lindkvist B: Pre-diagnostic levels of anionic trypsinogen, cationic trypsinogen, and pancreatic secretory trypsin inhibitor in relation to pancreatic cancer risk. Pancreatology 2010;10:229– 237.
- 39 Hirano T, Saluja A, Ramarao P, Lerch MM, Saluja M, Steer ML: Apical secretion of lysosomal enzymes in rabbit pancreas occurs via a secretagogue regulated pathway and is increased after pancreatic duct obstruction. J Clin Invest 1991;87:865–869.
- 40 Kereszturi E, Szmola R, Kukor Z, Simon P, Weiss FU, Lerch MM, Sahin-Tóth M: Hereditary pancreatitis caused by mutation-induced misfolding of human cationic trypsinogen: a novel disease mechanism. Hum Mutat 2009;30:575–582.
- 41 Lerch MM, Saluja AK, Dawra R, Saluja M, Steer ML: The effect of chloroquine administration on two experimental models of acute pancreatitis. Gastroenterology 1993; 104:1768–1779.
- 42 Sendler M, Dummer A, Weiss FU, Krüger B, Wartmann T, Scharffetter-Kochanek K, van Rooijen N, Malla SR, Aghdassi A, Halangk W, Lerch MM, Mayerle J: Tumour necrosis factor α secretion induces protease activation and acinar cell necrosis in acute experimental pancreatitis in mice. Gut 2013;62:430–439.
- 43 Gopinathan A, Denicola GM, Frese KK, Cook N, Karreth FA, Mayerle J, Lerch MM, Reinheckel T, Tuveson DA: Cathepsin B promotes the progression of pancreatic ductal adenocarcinoma in mice. Gut 2012;61:877–884.
- 44 Lerch MM, Lutz MP, Weidenbach H, Müller-Pillasch F, Gress TM, Leser J, Adler G: Dissociation and reassembly of adherens junctions during experimental acute pancreatitis. Gastroenterology 1997;113:1355–1366.
- 45 Mayerle J, Schnekenburger J, Krüger B, Kellermann J, Ruthenbürger M, Weiss FU, Nalli A, Domschke W, Lerch MM: Extracellular cleavage of E-cadherin by leukocyte elastase during acute experimental pancreatitis in rats. Gastroenterology 2005;129:1251–1267.
- 46 Schnekenburger J, Schick V, Krüger B, Manitz MP, Sorg C, Nacken W, Kerkhoff C, Kahlert A, Mayerle J, Domschke W, Lerch MM: The

calcium binding protein S100A9 is essential for pancreatic leukocyte infiltration and induces disruption of cell-cell contacts. J Cell Physiol 2008;216:558–567.

- 47 Michl P, Barth C, Buchholz M, Lerch MM, Rolke M, Holzmann KH, Menke A, Fensterer H, Giehl K, Löhr M, Leder G, Iwamura T, Adler G, Gress TM: Claudin-4 expression decreases invasiveness and metastatic potential of pancreatic cancer. Cancer Res 2003;63: 6265–6271.
- 48 Whitcomb DC, LaRusch J, Krasinskas AM, Klei L, Smith JP, Brand RE, Neoptolemos JP, Lerch MM, Tector M, Sandhu BS, Guda NM, Orlichenko L; Alzheimer's Disease Genetics Consortium, Alkaade S, Amann ST, Anderson MA, Baillie J, Banks PA, Conwell D, Coté GA, Cotton PB, DiSario J, Farrer LA, Forsmark CE, Johnstone M, Gardner TB, Gelrud A, Greenhalf W, Haines JL, Hartman DJ, Hawes RA, Lawrence C, Lewis M, Mayerle J, Mayeux R, Melhem NM, Money ME, Muniraj T, Papachristou GI, Pericak-Vance MA, Romagnuolo J, Schellenberg GD, Sherman S, Simon P, Singh VP, Slivka A, Stolz D, Sutton R, Weiss FU, Wilcox CM, Zarnescu NO, Wisniewski SR, O'Connell MR, Kienholz ML, Roeder K, Barmada MM, Yadav D, Devlin B: Common genetic variants in the CLDN2 and PRSS1-PRSS2 loci alter risk for alcohol-related and sporadic pancreatitis. Nat Genet 2012; 44:1349-1354.
- 49 Amundadottir L, Kraft P, Stolzenberg-Solomon RZ, Fuchs CS, Petersen GM, Arslan AA, Bueno-de-Mesquita HB, Gross M, Helzlsouer K, Jacobs EJ, LaCroix A, Zheng W, Albanes D, Bamlet W, Berg CD, Berrino F, Bingham S, Buring JE, Bracci PM, Canzian F, Clavel-Chapelon F, Clipp S, Cotterchio M, de Andrade M, Duell EJ, Fox JW Jr, Gallinger S, Gaziano JM, Giovannucci EL, Goggins M, González CA, Hallmans G, Hankinson SE, Hassan M, Holly EA, Hunter DJ, Hutchinson A, Jackson R, Jacobs KB, Jenab M, Kaaks R, Klein AP, Kooperberg C, Kurtz RC, Li D, Lynch SM, Mandelson M, McWilliams RR, Mendelsohn JB, Michaud DS, Olson SH, Overvad K, Patel AV, Peeters PH, Rajkovic A, Riboli E, Risch HA, Shu XO, Thomas G, To-

bias GS, Trichopoulos D, Van Den Eeden SK, Virtamo J, Wactawski-Wende J, Wolpin BM, Yu H, Yu K, Zeleniuch-Jacquotte A, Chanock SJ, Hartge P, Hoover RN: Genome-wide association study identifies variants in the ABO locus associated with susceptibility to pancreatic cancer. Nat Genet 2009;41:986–990.

- 50 Weiss FU, Schurmann C, Guenther A, Ernst F, Teumer A, Mayerle J, Simon P, Völzke H, Radke D, Greinacher A, Kuehn JP, Zenker M, Völker U, Homuth G, Lerch MM: Fucosyltransferase 2 (FUT2) non-secretor status and blood group B are associated with elevated serum lipase activity in asymptomatic subjects, and an increased risk for chronic pancreatitis: a genetic association study. Gut 2015;64:646–656.
- 51 Menges M, Lerch MM, Zeitz M: The double duct sign in patients with malignant and benign pancreatic lesions. Gastrointest Endosc 2000;52:74–77.
- 52 Gudjonsson B: Cancer of the pancreas. 50 years of surgery. Cancer 1987;60:2284–2303.
- 53 Neoptolemos JP, Stocken DD, Friess H, Bassi C, Dunn JA, Hickey H, Beger H, Fernandez-Cruz L, Dervenis C, Lacaine F, Falconi M, Pederzoli P, Pap A, Spooner D, Kerr DJ, Büchler MW: European Study Group for Pancreatic Cancer. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer. N Engl J Med 2004;350: 1200–1210.
- 54 Burris HA 3rd, Moore MJ, Andersen J, Green MR, Rothenberg ML, Modiano MR, Cripps MC, Portenoy RK, Storniolo AM, Tarassoff P, Nelson R, Dorr FA, Stephens CD, Von Hoff DD: Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. J Clin Oncol 1997;15:2403– 2413.
- 55 Cunningham D, Chau I, Stocken DD, Valle JW, Smith D, Steward W, Harper PG, Dunn J, Tudur-Smith C, West J, Falk S, Crellin A, Adab F, Thompson J, Leonard P, Ostrowski J, Eatock M, Scheithauer W, Herrmann R, Neoptolemos JP: Phase III randomized comparison of gemcitabine versus gemcitabine plus capecitabine in patients with advanced pancreatic cancer. J Clin Oncol 2009;27:5513–5518.

Development of Pancreatic Cancer: Targets for Early Detection and Treatment