

Molecular insight into the IRE1 α -mediated type I interferon response induced by proteasome impairment in myeloid cells of the brain.

Supplementary Material

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Figure S1. Transcriptional regulation of proteasome genes in primary microglia with proteasome impairment. (A) Quantitative RT-PCR analysis of *Psmb8*, *Psmb9* and *Psmb10* mRNA levels encoding immunoproteasome catalytic subunits as well as *Psm2* and *Psm6* mRNA levels encoding standard proteasome catalytic subunits in primary microglia treated for 8 hours either with 200 nM of ONX-0914 or 50 nM of bortezomib. Mean \pm SEM (n=3); *: P < 0.05, **: P < 0.01, ***: P < 0.001, unpaired two-sided t-test. (B) Immunoblotting depicting expression of NRF1 after bortezomib treatment.

Figure S2. Proteasome activity in BV-2 cells treated with different doses of bortezomib. (A) Viability of BV-2 cells in response to different doses of bortezomib. Cells were treated with the indicated doses of BTZ for 6 hours and analyzed with the PreMix WST-1 Cell Proliferation Assay System. (B) Immunoblotting depicting expression of UPR drivers in response to indicated doses of bortezomib. (C) Presence of a spliced variant of XBP1s mRNA in response to bortezomib treatment was detected by PCR. XBP1u was observed as a 204-bp band, and XBP1s was observed as a 178-bp band. Quantitative RT-PCR analysis of (D) *Ifn β 1*, (E) *Cxcl-10*, *Ddit3*/CHOP, *Eif2ak2*/PKR and (F) *Il-6* mRNA levels. Mean \pm SEM (n=3 technical replicates).

Figure S3. Proteasome activity in BV-2 cells treated with 50 nM bortezomib. (A) Cytotoxicity test in response to 50 nM of bortezomib. Cells were treated for the indicated period of time with 50 nM of bortezomib and analyzed with the PreMix WST-1 Cell Proliferation Assay System. (B) An in-gel overlay experiment depicting chymotryptic-like activity of native proteasome complexes before and after the treatment with 50 nM of bortezomib. (C) Quantitative analysis of the chymotryptic-like activity of proteasomes treated with 50 nM of bortezomib. Measurement was performed on native whole BV-2 cell lysates. (D) An in-gel analysis of proteasome activity after bortezomib treatment by use of the active site probe UbiQ-018 that fluorescently and covalently labels the β subunits of the proteasome in native whole cell lysates of BV-2. (E) Western blot analysis of poly-ubiquitin conjugates in BV-2 cells treated with bortezomib. (F) Quantification of ROS formation in cells treated with bortezomib. ROS was labelled with 2'-7'-dichlorofluorescein diacetate (DCFDA). Mean \pm SEM (n=2)

Figure S4. Transcriptional regulation of proteasome genes in BV-2 cells following treatment with bortezomib. (A) Quantitative RT-PCR analysis of immunoproteasome subunit *Psmb8*, *Psmb9* and *Psmb10* mRNA levels encoding immunoproteasome catalytic

subunits as well as *Pσμα2* and *Pσmb6* mRNA levels encoding standard proteasome catalytic subunits in BV-2 treated with 50 nM of bortezomib for indicated time. Mean \pm SEM (n=5); *: P < 0.05, ***: P < 0.001, ****: P < 0.0001, unpaired two-sided t-test. **(B)** Immunoblotting depicting expression of NRF1 in BV-2 cells treated with bortezomib. **(C)** IL-6 expression levels in BV-2 upon bortezomib treatment measured by ELISA.

Figure S5. Analysis of UPR and type I IFN response induction in BV-2 cells treated with ONX-0914. **(A)** Viability of BV-2 cells in response to different doses of ONX-0914. Cells were treated with the indicated doses of ONX-0914 for 6 hours and analyzed with the PreMix WST-1 Cell Proliferation Assay System. **(B)** Immunoblotting depicting expression of UPR drivers in response to indicated doses of ONX-0914. **(D)** Cytotoxicity test in response to 200 nM of ONX-0914. Cells were treated with the indicated dose of ONX-0914 up to 24 hours and analyzed with the PreMix WST-1 Cell Proliferation Assay System. **(D)** Expression of standard- and immunoproteasome catalytic subunits and as well as UPR proteins visualized by western blotting. **(E)** Examination of the XBP1s mRNA splicing, visualized by PCR. **(F)** Immunoblotting depicting expression of type I IFN response drivers in BV-2 cells treated with ONX-0914.

Figure S6. Analysis of UPR and type I IFN response induction in BV-2 cells treated with IFN β 1. BV-2 cells were treated with 100U of IFN β 1 up to 10 hours. Expression of **(A)** key drivers of type I IFN response, **(B)** standard- and immunoproteasome catalytic subunits and **(C)** CHOP and ATF6 proteins was visualized by immunoblotting.

Figure S7. Analysis of effects of IFN α/β receptor blocker on the expression of interferon-stimulated genes in response to bortezomib treatment. **(A)** Immunoblotting depicting expression of type I IFN response drivers in BV-2 cells in response to a treatment with IFNAR blocking antibody (10 μ g/ml) or the isotype IgG control. Quantitative RT-PCR analysis of **(B)** *Ifn β 1*, **(C)** *Il-6*, **(D)** *Cxcl-10* and *Eif2ak2/ PKR* mRNA levels from whole RNA extracts of BV-2 cells. Mean \pm SEM (n=3 technical replicates). **(E)** Immunoblotting depicting expression of main UPR proteins in BV-2 cells in response to the treatment with IFNAR blocking antibody (10 μ g/ml) or the isotype IgG control.

Figure S8. Induction of Interferon Stimulated Genes (ISGs) in BV-2 cells treated with IFN β 1. Quantitative RT-PCR analysis of ISGs mRNA levels in BV-2 cells treated with 100U of interferon β 1 for indicated period of time. Mean \pm SEM (n=3); *: P < 0.05, **: P < 0.01, unpaired two-sided t-test.

Figure S9. Induction of Interferon Stimulated Genes (ISGs) in BV-2 cells following treatment with ONX-014. Quantitative RT-PCR analysis of ISGs mRNA levels in BV-2 cells. Mean \pm SEM.

Figure S10. Proteasome subunits expression after the interference with the endonuclease activity of IRE1 α . (A) Cytotoxicity test in response to proteasome and IRE1 α inhibition with to 50 nM of bortezomib and 100 μ M of 4 μ 8C inhibitor or the diluent, DMSO (0.1% v/v), respectively. Cell viability was analyzed with the PreMix WST-1 Cell Proliferation Assay System. (B) Immunoblotting depicting expression of proteasome subunits in BV-2 cells treated with 50 nM of bortezomib. Cells were pre-treated for 2 hours with 100 μ M of the 4 μ 8C inhibitor or with DMSO as a control. (C) Quantitative RT-PCR analysis of *Ddit3/CHOP* and *Eif2ak2/ PKR* mRNA levels in BV-2 cells after the IRE1 inhibition. Mean \pm SEM (n=3). (D) Viability of THP-1 cells in response to different doses of bortezomib. Cells were treated for 6 hours and analyzed using the PreMix WST-1 Cell Proliferation Assay System. (E) Cytotoxicity test of THP-1 cells in response to 100 nM of bortezomib. Cells were treated with the indicated dose of BTZ up to 10 hours and analyzed using the PreMix WST-1 Cell Proliferation Assay System. (F) Immunoblotting depicting expression of type I IFN response drivers in THP1 cells treated with IRE1 α inhibitor and 100 nM of bortezomib.

Figure S11. Proteasome subunits expression after the interference with TBK1 kinase activity. (A) Cytotoxicity test in response to proteasome and TBK1 inhibition with to 50 nM of bortezomib and 100 μ M of BX975 inhibitor or the diluent, DMSO (0.1% v/v), respectively. Cell viability was analyzed with the PreMix WST-1 Cell Proliferation Assay System. (B) Immunoblotting depicting expression of proteasome subunits in BV-2 cells treated with 50 nM of bortezomib. Cells were pre-treated for 1 hour with 1 μ M of the TBK1 inhibitor – BX795 or with DMSO as a control. (C) Quantitative RT-PCR analysis of *Eif2ak2/ PKR* mRNA level in BV-2 cells after TBK1 inhibition. Mean \pm SEM (n=3).

Figure S12. Induction of Interferon Stimulated Genes (ISGs) in CHOP deficient BV-2 cells following bortezomib treatment. (A) Cytotoxicity test in response to proteasome inhibition and/ or CHOP knockdown. Cell viability was analyzed with the PreMix WST-1 Cell Proliferation Assay System. Quantitative RT-PCR analysis of (B) *Ifn β 1* mRNA level and (C) *Cxcl10* as well as *Eif2ak2/PKR* mRNA levels in BV-2 cells. Mean \pm SEM (n=3).

Table S1. List of primers used for RT-qPCR analysis.

Target gene	Sequence
RPLP0	forward 5'-CACACTCCATCATCAATGGG-3' reverse 5'-TAGTCGAAGAGACCGAATCC-3'
IFN β 1	forward 5'-TGGGAGATGTCCTCAACTGC-3' reverse 5'-TGCAACCACCACTCATTCTGA-3'
CXCL-10	forward 5'-GTGTTGAGATCATTGCCACG-3' reverse 5'-AAGGAGCCCTTTAGACCTT-3'
ISG-15	forward 5'-GAAGCAGACTCCTTAATTCCA-3' reverse 5'-CTTTAGGTCCCAGGCCATTG-3'
EIF2AK2/ PKR	forward 5'-ACAAATCGTGACCGGAGTGG-3' reverse 5'-CAGGTCCGTCCTTGGGTTTC-3'
USP18	forward 5'-ATTGAAGAGGATAACAGTGCC-3' reverse 5'-CGTGATCTGGTCCTTAGTCA-3'
PSMB8	forward 5'-TTGGTGGTGACCAGGAAAGG-3' reverse 5'-GAGAGCCGAGTCCCATTGTC-3'
PSMB9	forward 5'-GAAGAAGTCCACACCGGGA-3' reverse 5'-GGTTCATATACCTGTCCCCC-3'
PSMB10	forward 5'-ACAAAAGCTGCGAGAAGAT-3' reverse 5'-TGTGATGGCTTCCACCAA-3'
PSMA2	forward 5'-GTTCGTCCATTTGGTGTTTC-3' reverse 5'-AGGGTTAAGATGGCTGTATGA-3'
PSMB6	forward 5'-CAGCTTGGTTCCACAGTATT-3' reverse 5'-TGTCTTACCATCATACCCC-3'
CHOP	forward 5'-GAACCTGAGGAGAGAGTGTT-3' reverse 5'-TATAGGTGCCCCAATTTCA-3'
IL-6	forward 5'-ACCAAGAGATAAGCTGGAGT-3' reverse 5'-TAGGCATAACGCACTAGGTT-3'

Figure S1

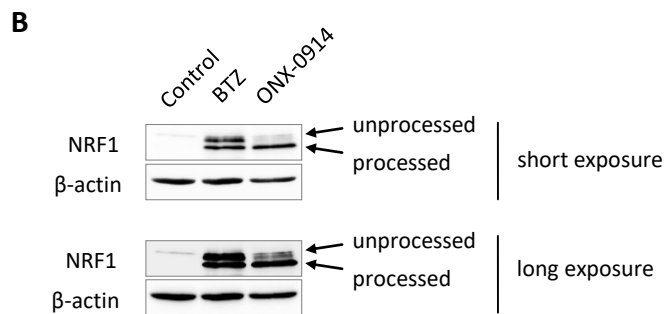
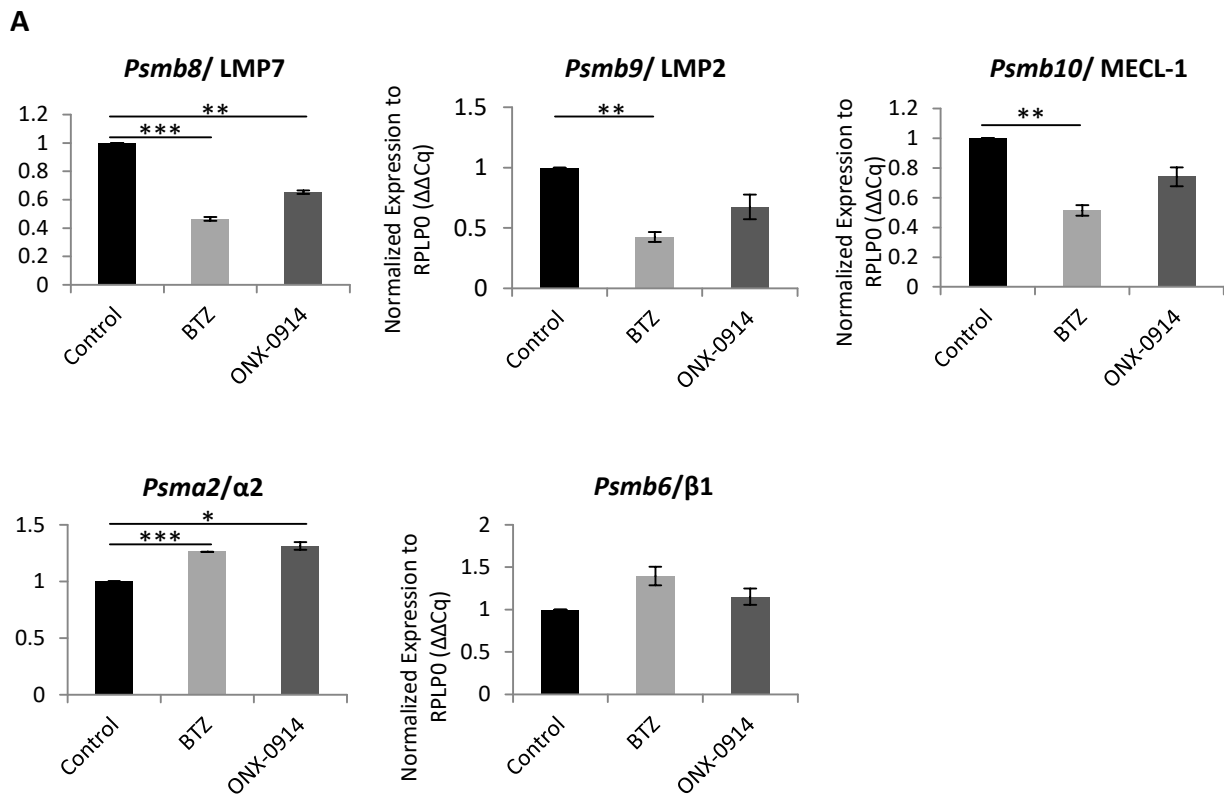


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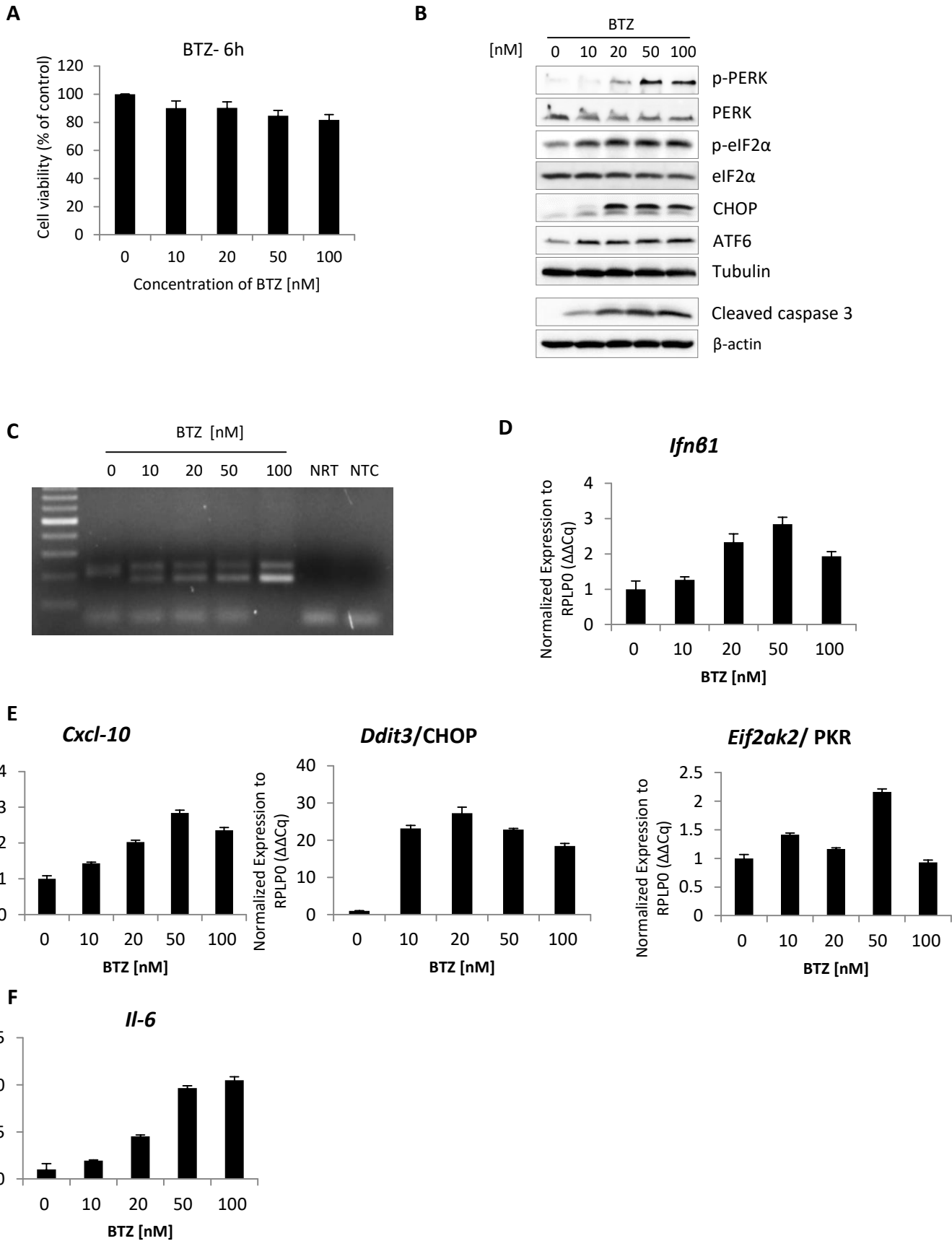


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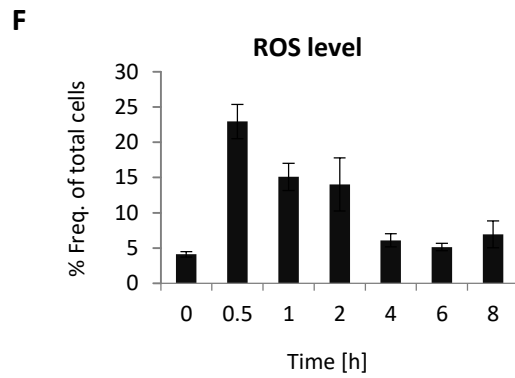
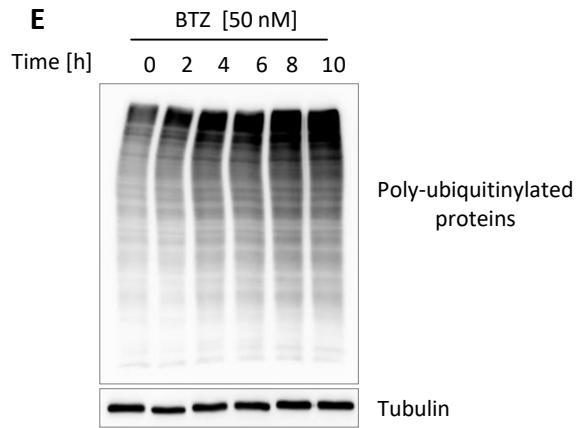
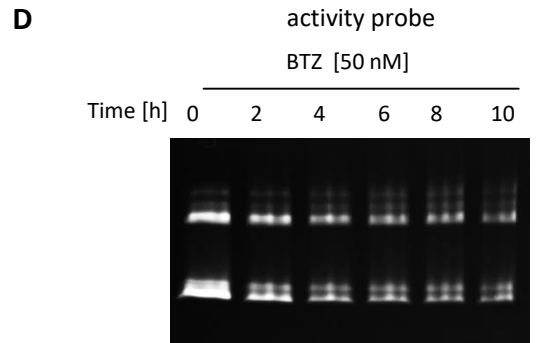
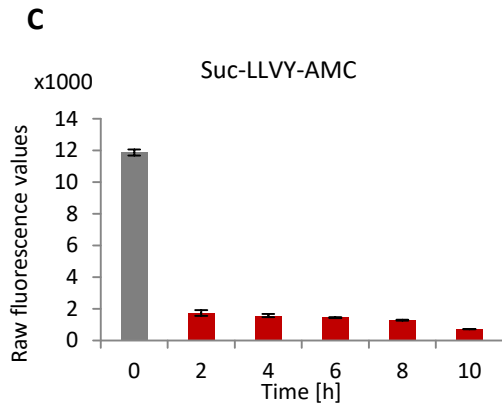
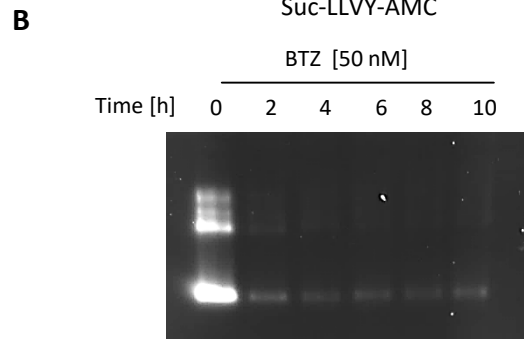
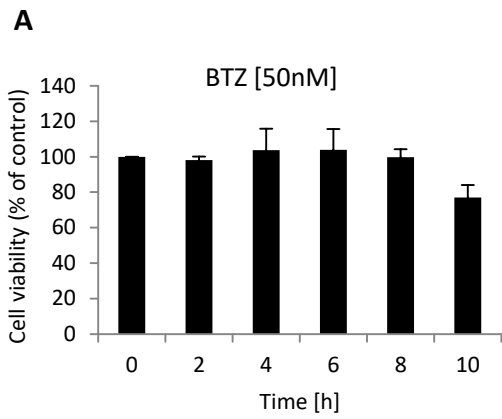
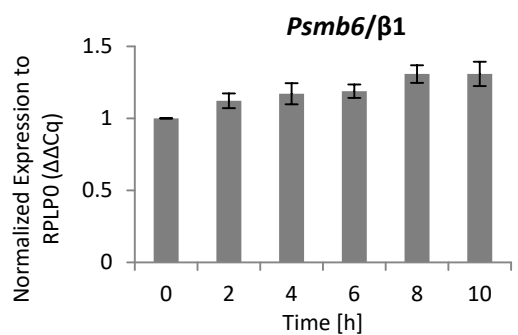
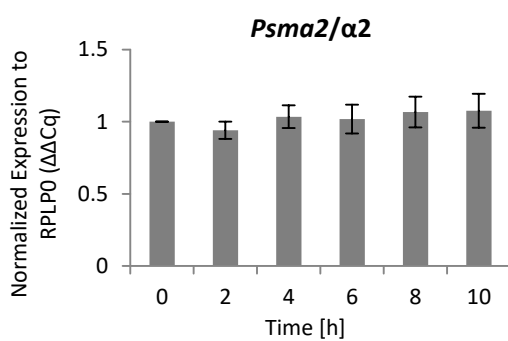
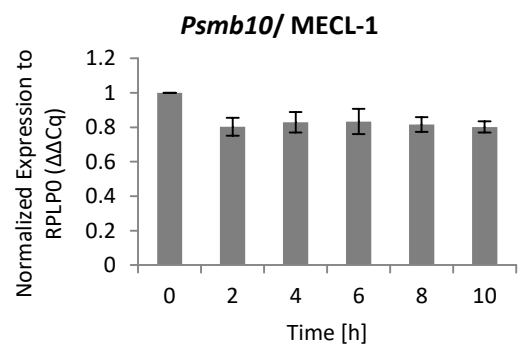
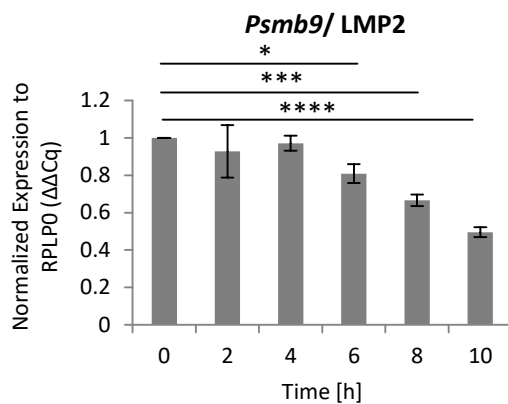
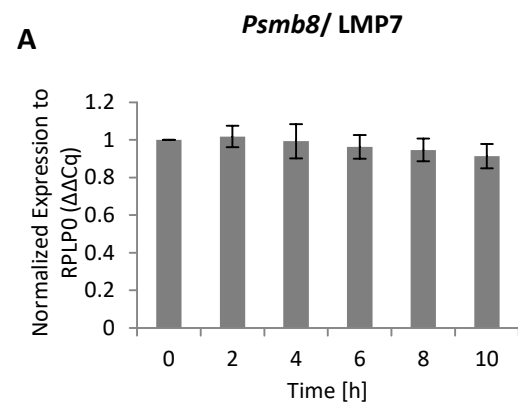
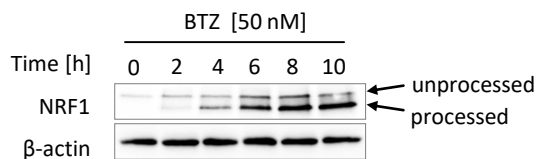


Figure S4

A



B



C

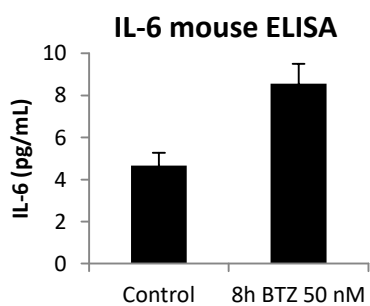


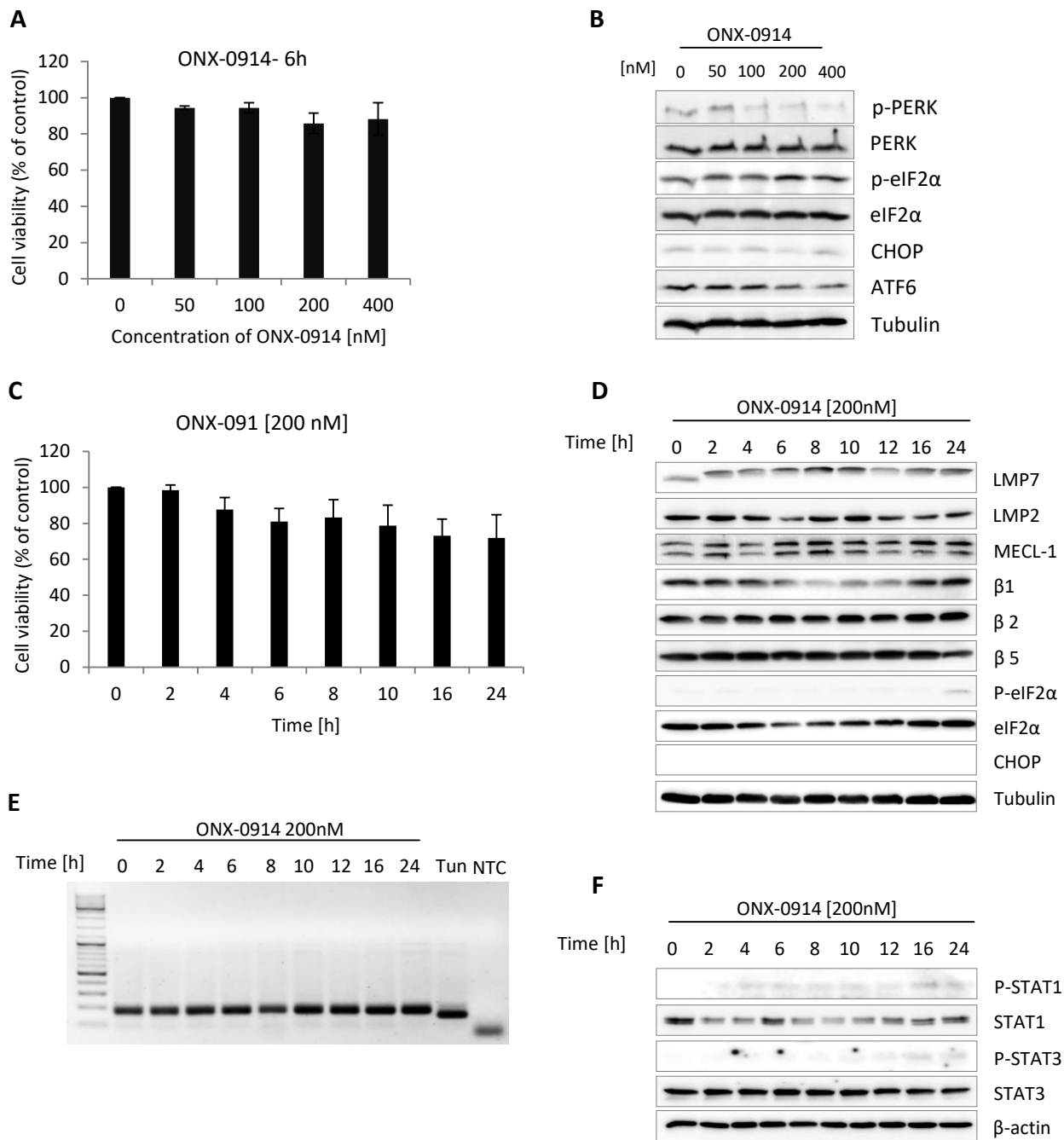
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Figure S6

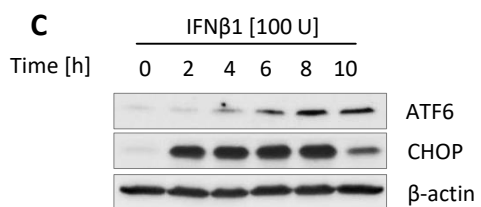
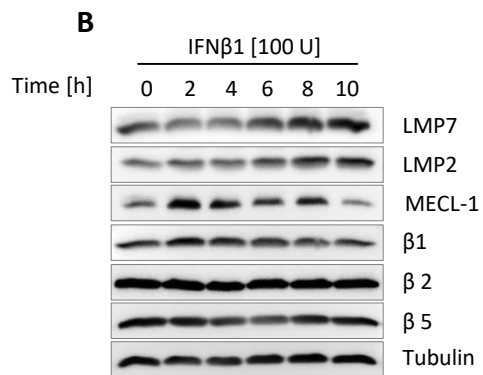
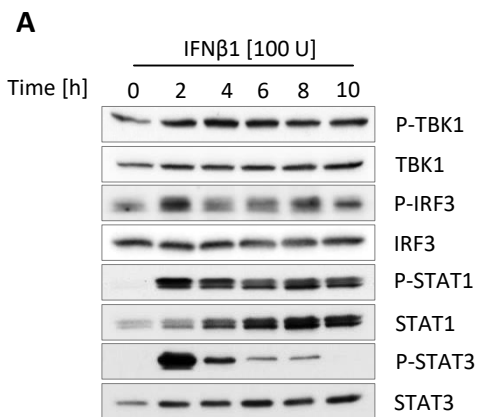


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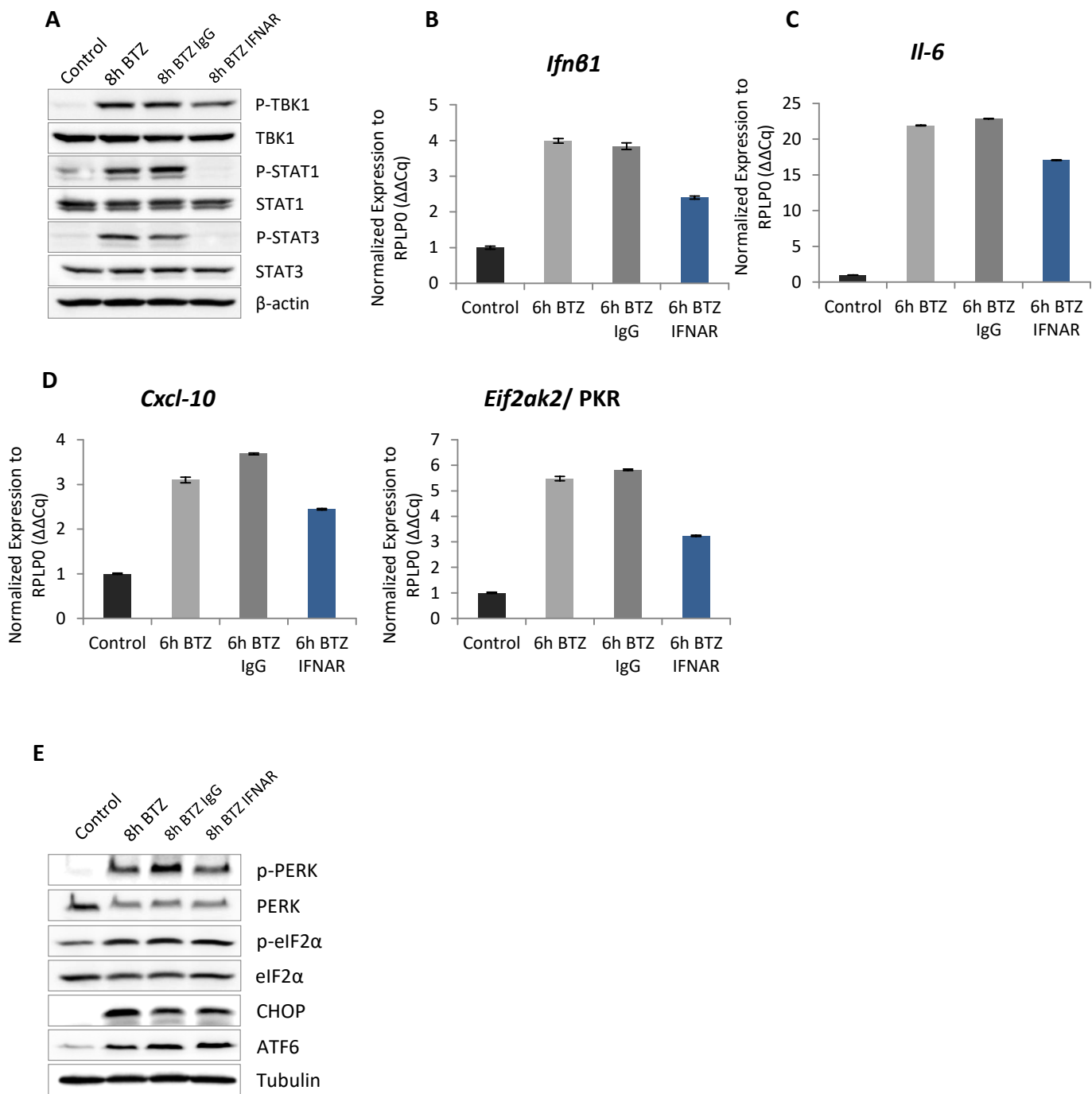


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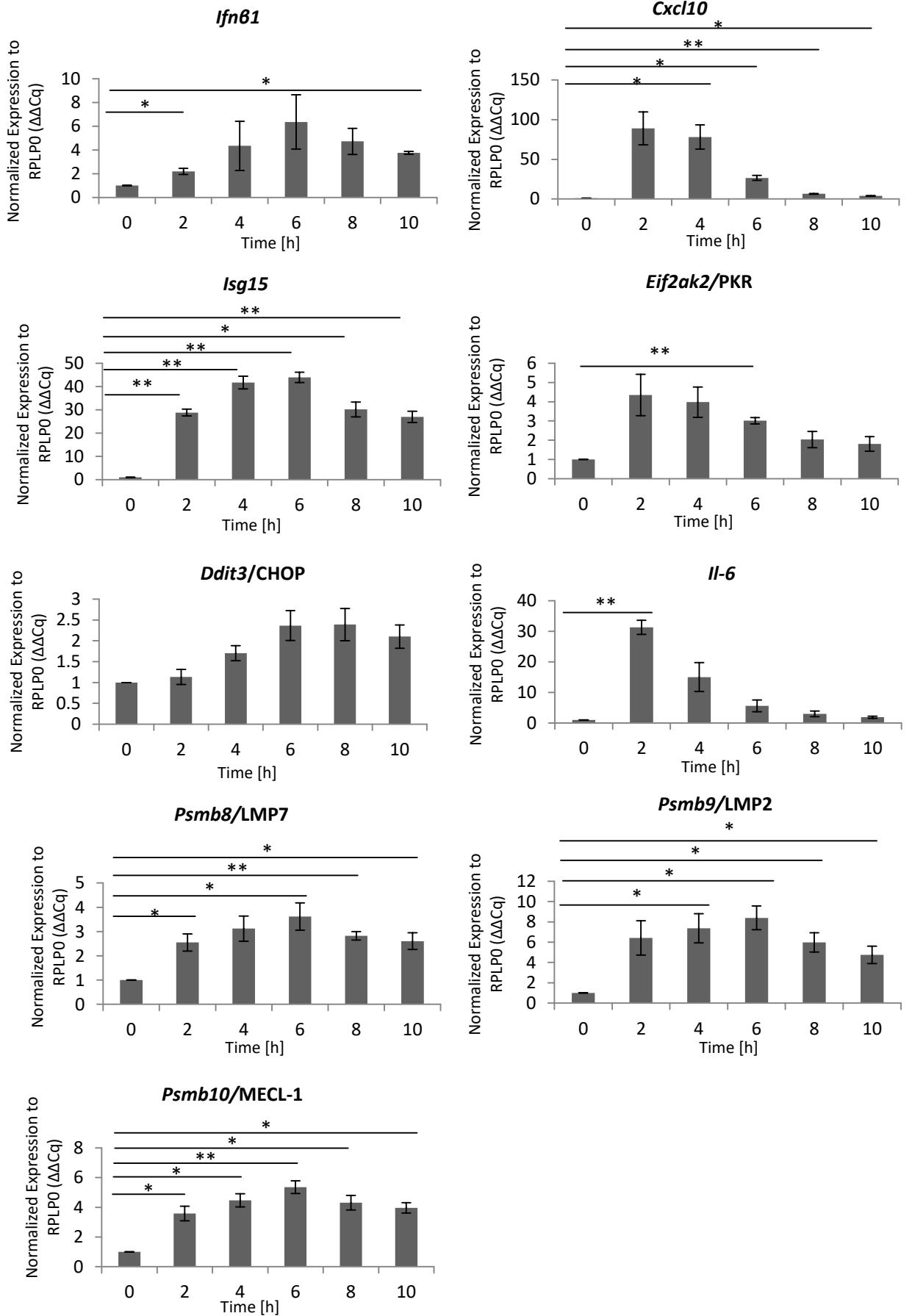


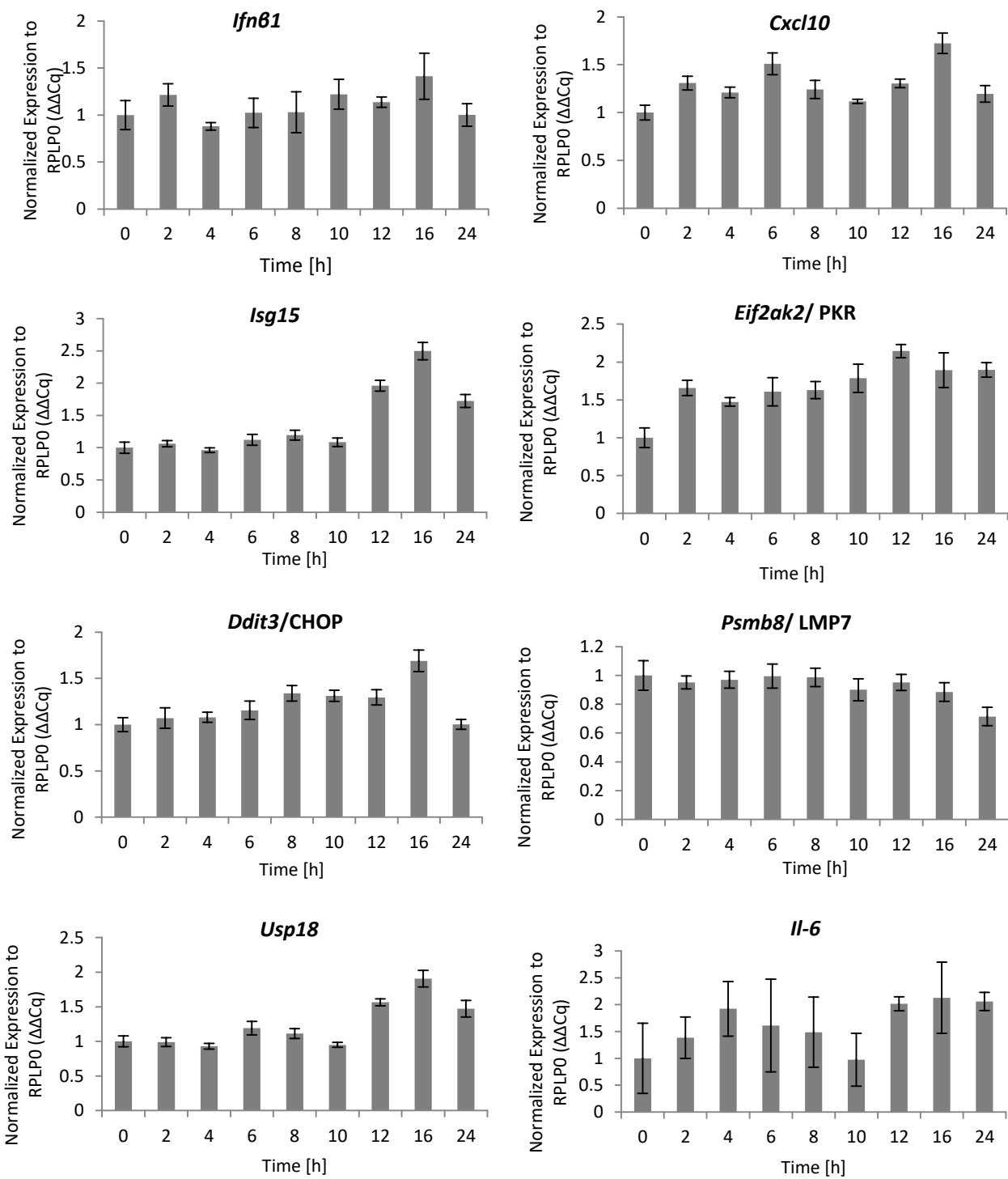
Figure S9

Figure S10

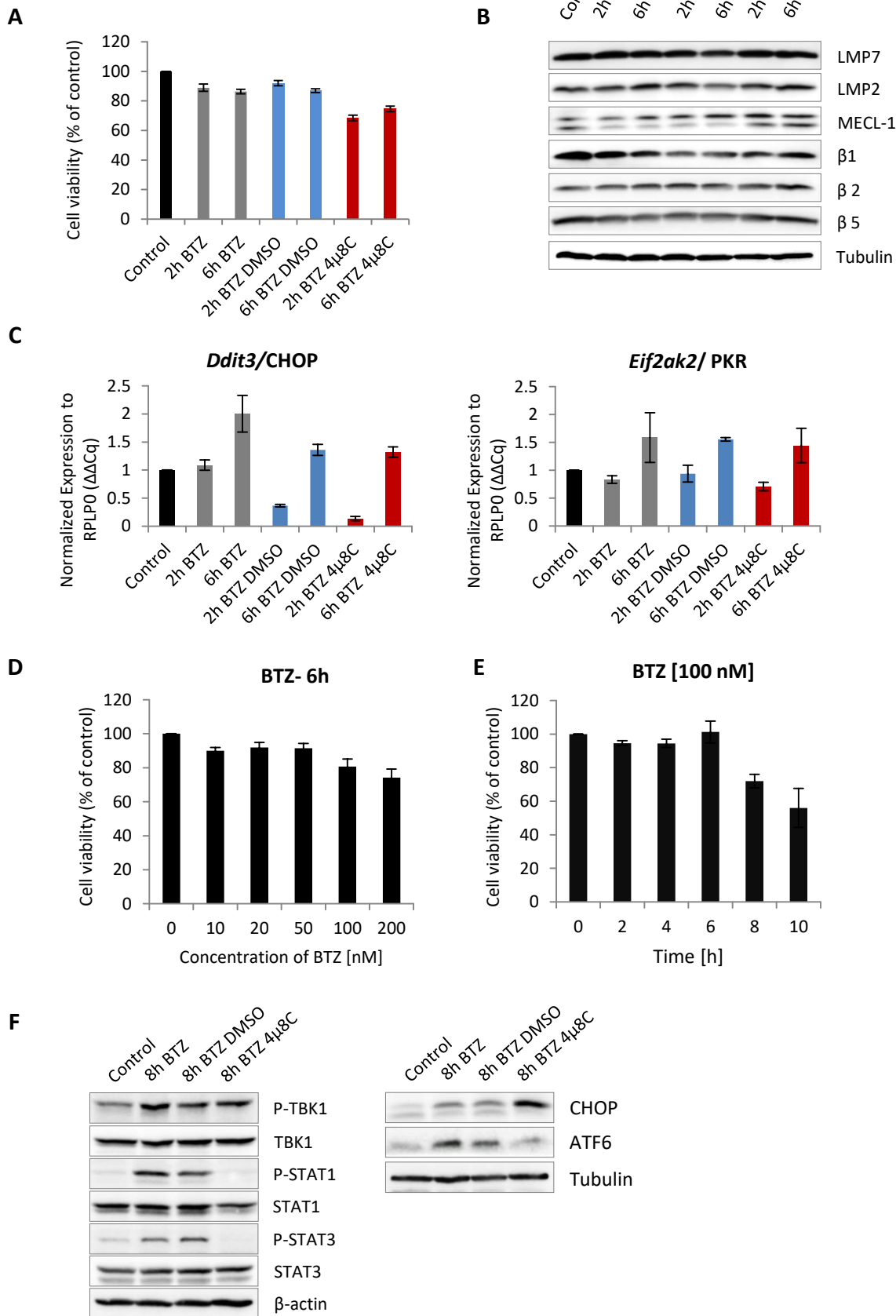
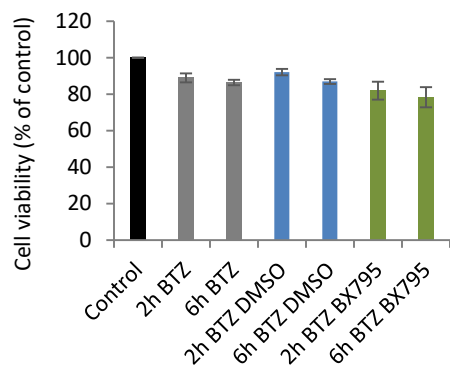
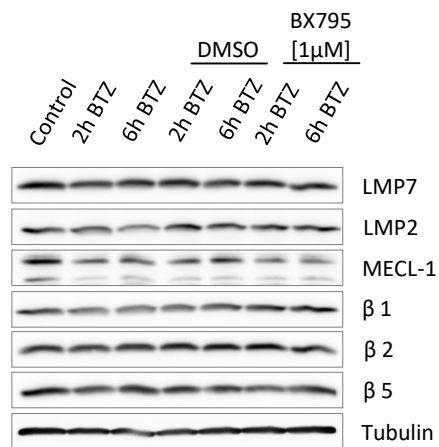


Figure S11

A



B



C

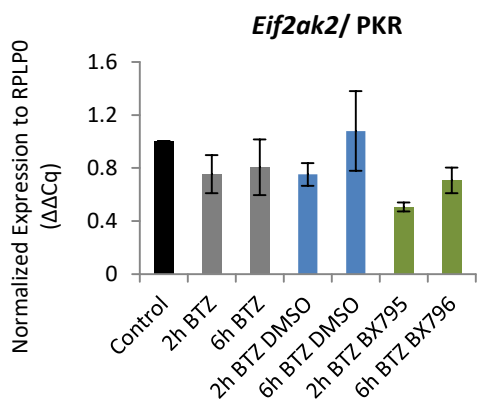
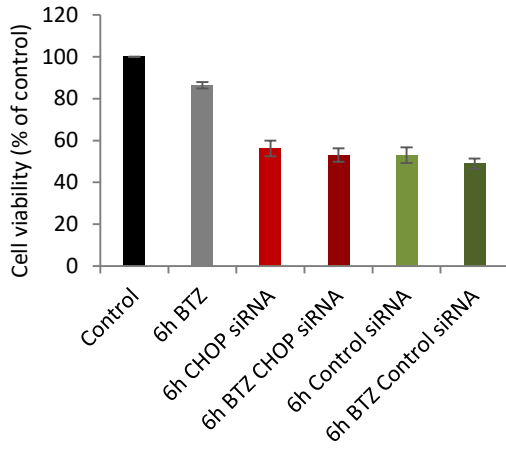
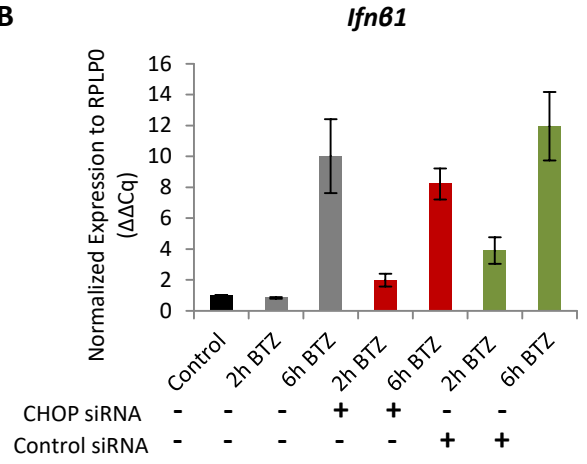


Figure S12

A



B



C

