

Genotype-environment interactions rule the response of a widespread butterfly to temperature variation

Franziska Günter¹  | Michaël Beaulieu¹  | Kasimir F. Freiberg¹ | Ines Welzel¹ |
Nia Toshkova² | Anamarija Žagar³ | Tatjana Simčič³ | Klaus Fischer¹ 

¹Zoological Institute and Museum, Greifswald University, Greifswald, Germany

²AgroBio Institute, Sofia, Bulgaria

³National Institute of Biology, Ljubljana, Slovenia

Correspondence

Franziska Günter, Zoological Institute and Museum, Greifswald University, Soldmannstraße 14, D-17489 Greifswald, Germany.
Email: Franziska.guenter@uni-greifswald.de

Present address

Klaus Fischer, Institute for Integrated Natural Sciences, University Koblenz-Landau, Koblenz, D-56070, Germany

Funding information

Deutsche Forschungsgemeinschaft, Grant/Award Number: RESPONSE (DFG GRK 2010)

Abstract

Understanding how organisms adapt to complex environments is a central goal of evolutionary biology and ecology. This issue is of special interest in the current era of rapidly changing climatic conditions. Here, we investigate clinal variation and plastic responses in life history, morphology and physiology in the butterfly *Pieris napi* along a pan-European gradient by exposing butterflies raised in captivity to different temperatures. We found clinal variation in body size, growth rates and concomitant development time, wing aspect ratio, wing melanization and heat tolerance. Individuals from warmer environments were more heat-tolerant and had less melanised wings and a shorter development, but still they were larger than individuals from cooler environments. These findings suggest selection for rapid growth in the warmth and for wing melanization in the cold, and thus fine-tuned genetic adaptation to local climates. Irrespective of the origin of butterflies, the effects of higher developmental temperature were largely as expected, speeding up development; reducing body size, potential metabolic activity and wing melanization; while increasing heat tolerance. At least in part, these patterns likely reflect adaptive phenotypic plasticity. In summary, our study revealed pronounced plastic and genetic responses, which may indicate high adaptive capacities in our study organism. Whether this may help such species, though, to deal with current climate change needs further investigation, as clinal patterns have typically evolved over long periods.

KEYWORDS

climatic adaptation, cline, environmental gradient, genetic adaptation, local adaptation, *Pieris napi*, thermal melanisation, thermoregulation

Data deposited at Dryad: <https://doi.org/10.5061/dryad.h9w0vt4dw>

The peer review history for this article is available at <https://publons.com/publon/10.1111/jeb.13623>

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Journal of Evolutionary Biology* published by John Wiley & Sons Ltd on behalf of European Society for Evolutionary Biology

1 | INTRODUCTION

Understanding how organisms adapt to complex environments is of special interest in the current era of rapidly changing climatic conditions (Parmesan, 2006; Parmesan et al., 1999). Widespread species have to cope with a variety of environmental conditions within their distribution ranges. Accordingly, selective pressures show spatial variation, which often results in geographic differences in fitness-related traits (Kawecki & Ebert, 2004). Evidence for such local adaptation stems, among others, from clinal variation, suggesting a contribution of selection to the differentiation among populations (Bradshaw & Holzapfel, 2008; Chown & Klok, 2003; Robinson & Partridge, 2001; Sambucetti, Loeschcke, & Norry, 2006; Van Doorslaer & Stoks, 2005). Putative examples include increased body size (Arnett & Gotelli, 1999a; Chown & Klok, 2003) and growth rates (Van Doorslaer & Stoks, 2005) but reduced developmental time (Robinson & Partridge, 2001) and heat tolerance (Hoffmann & Rieseberg, 2008; Kapun, Fabian, Goudet, & Flatt, 2016) with increasing latitude.

As temperature is an important ecological factor, thermal selection is believed to cause or at least contribute to local adaptation (Reeve, Fowler, & Partridge, 2000). Potential target traits of thermal selection include body size, locomotory performance and coloration. Animals are generally predicted to be larger in cooler environments based on both the Bergmann's rule and the temperature-size rule (Arnett & Gotelli, 1999b; Chown & Gaston, 1999). However, converse Bergmann's clines have also been found in insects (Blanckenhorn & Demont, 2004; Blanckenhorn, Stillwell, Young, Fox, & Ashton, 2006). Due to a shorter season length and lower ambient temperatures, there is also strong selection on fast growth at higher latitudes. Such countergradient variation helps animals to maintain their optimal body size even under challenging conditions (Arendt, 1997; Blanckenhorn & Demont, 2004). Likewise, in flying insects increased flight performance is expected in cooler environments. This is because high body temperature is needed for flight, resulting in thermal limitations in the cold (Berwaerts, Dyck, & Aerts, 2002; Dahlggaard, Hasson, & Loeschcke, 2001; Norry, Bublly, & Loeschcke, 2001). Moreover, thermal melanization increasing heat absorption has been frequently found in cooler environments (Ellers & Boggs, 2004; Watt, 1968).

Although living in cool environments may constrain life histories (see above), high temperatures may also involve physiological costs. They have direct effects on cellular processes and metabolism, and may induce oxidative stress and concomitantly costly antioxidant responses (Gillooly, Brown, West, Savage, & Charnov, 2001). Oxidative stress is an imbalance between the production of reactive oxygen species (ROS), typically increasing with metabolism and thus temperature in ectotherms, and antioxidant defences, resulting in higher oxidative damage (Ju, Wei, Wang, Zhou, & Li, 2014; Tumminello & Fuller-Espie, 2013). Important parameters in this context are consequently metabolic rate potentially affecting the production of ROS, the degree of oxidative damage and antioxidant defence mechanisms (Ju et al., 2014; Žagar, Carretero, Marguč, Simčič, & Vrezec, 2018).

In an earlier study using exclusively field-collected individuals of the butterfly *Pieris napi*, we found clinal variation in body size and wing coloration (Günter et al., 2019). Specifically, we showed that body size and the wings' yellow reflectance decreased whereas wing melanization increased with increasing latitude and altitude. However, the sources of variation, namely the relative contribution of genetic versus plastic responses, remained unknown in this earlier study and are a scientific field that still has open questions and needs more investigation (for a review, see Crispo, 2008). Against this background, we here set out to investigate clinal variation in *P. napi* using a common garden design with replicated populations from Italy to Sweden. We investigate traits related to body size, flight performance, wing coloration, physiological status and stress resistance, as local adaptation in such traits is at least partly still not well understood. We hypothesize that genetic variation in such traits is widespread as is plasticity therein and that responses are modulated by interactions between both sources of variation. Specifically, we predict (a) reduced growth rates (countergradient variation), metabolic potential, flight performance and wing melanization but increased heat tolerance and defence mechanisms to fight oxidative stress at lower latitudes (genetic variation), and (b) higher growth rates, metabolic potential, heat tolerance and defence mechanisms to fight oxidative stress but reduced body size, flight performance and wing melanization at a higher compared with a lower developmental temperature (plastic responses).

2 | MATERIALS AND METHODS

2.1 | Study organism and experimental populations

The green-veined white butterfly *Pieris napi* L. is a widespread temperate-zone butterfly, occurring throughout Europe and the temperate zone of Asia (Ebert & Rennwald, 1993). In Europe, it is one of the most common butterflies. Nevertheless, it is predicted to suffer from anthropogenic climate change because of its association with moist habitats (Fox et al., 2015). Larvae feed on a variety of Brassicaceae, with *Alliaria petiolata* Cavara & Grande being the most important one. Brassicaceae are also the preferred nectar plants. *Pieris napi* is a polyandrous species in which males transfer large nuptial gifts to their female partners (Bergström & Wiklund, 2002). Accordingly, males are larger than females (Wiklund & Kaitala, 1995).

2.2 | Sampling locations

For this study, we collected freshly eclosed, spring generation females from a total of nine populations along a latitudinal gradient from northern Italy to Sweden (Figure 1). We sampled three replicate populations each in northern Italy (I: Torino 45.11°N/7.48°E, Pavia 45.21°N/9.27°E and Mantova 45.21°N/10.75°E), northern Germany (G: Wahrenholz 52.64°N/10.61°E, Rathenow

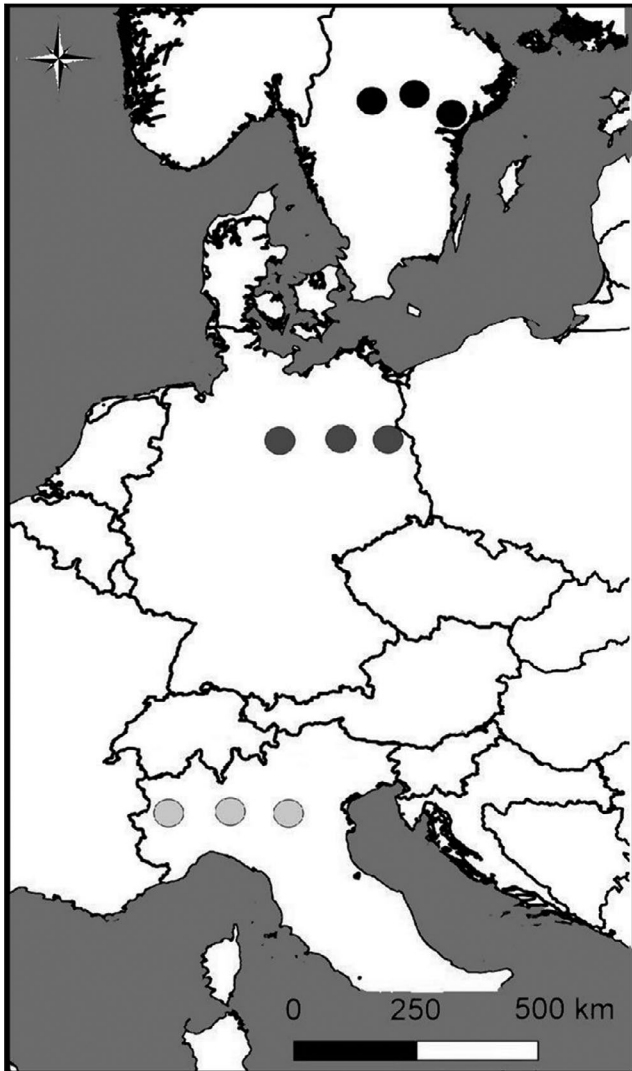


FIGURE 1 Map of sampling locations of *Pieris napi* individuals used in the present study. Italy (light grey circles), Germany (dark grey circles) and Sweden (black circles)

52.65°N/12.44°E and Strausberg 52.60°N/13.86°E) and Sweden (S: Örebro 59.29°N/15.01°E, Eskilstuna 59.36°N/16.54°E and Stockholm 58.95°N/17.58°E). Mean annual (Italy: 13°C; Germany: 9°C; and Sweden: 6°C) and the mean temperature during the vegetation period (May to September; Italy: 21°C; Germany: 16°C; and Sweden: 14°C) followed a temperature gradient across latitudes, whereas precipitation is higher in Italy than in Germany and Sweden (Table S1). The minimal straight distance between two populations was 73 km, and the total latitudinal gradient spanned ca. 1,660 km. We collected a total of 74 females from Italy, 94 from Germany and 76 from Sweden between 19 April and 14 June 2016. All females were afterwards transferred to Greifswald University for egg laying. Of these, 36, 37 and 37 females, respectively, produced sufficient offspring and thus contributed to the experiments below, resulting in a total number of 1,829 offspring.

2.3 | Experimental design

Field-caught females were kept individually in translucent 1-litre plastic pots covered with gauze, which were placed into a climate chamber set at a constant temperature of 25°C, 65% relative humidity and a photoperiod of L18:D6. Females were fed ad libitum with water, a 10% sugar solution and additionally flowers (e.g. *Sambucus nigra*, *Taraxacum spec.* and *Senecio spec.*). For oviposition, they were provided daily with a fresh cutting of *A. petiolata*. The resulting eggs were collected daily, counted and kept separated by oviposition day and female under the above conditions until hatching. Eggs were collected until the death of the females. After hatching, all larvae were transferred individually to translucent plastic pots (250 ml) lined with moist tissue and cuttings of *A. petiolata* ad libitum. Host plants were replaced on a daily basis. On day 3 after hatching, larvae were randomly divided among two rearing temperatures (18 and 25°C), using a split-brood design. These temperatures were chosen to mimic summer conditions (July) in Germany/Sweden and Italy (Table S1). Larval rearing took place in climate cabinets (Sanyo MLR-351H) at 60% relative humidity and L18:D6. For each individual, we recorded larval development time, pupal development time and pupal mass (one day after pupation, KERN ABJ-120-4M; 0.1 mg accuracy). Larval growth rate was calculated as the natural logarithm of pupal mass/larval time. One day after adult eclosion, butterflies were subjected to a heat knock-down assay. Therefore, they were individually placed into translucent plastic pots (100 ml) in a randomized block design and exposed to a constant temperature of 43°C (climate cabinet Sanyo MIR-553). The time until physical knock-down, characterized by an inability to move in a coordinated manner, was recorded. Butterflies were afterwards frozen at -80°C for later analyses.

Frozen butterflies were later thawed, and their adult fresh mass was measured with a digital scale (KERN ABJ-120-4M). Then, the head, wings and legs were removed, and the thorax and abdomen were separated before being weighed. The thorax-abdomen ratio was calculated as the thorax mass divided by the abdomen mass. Butterfly wings were used to measure wing morphology and melanization. Therefore, we took a photograph of one dorsal forewing and one ventral hindwing per individual under standardized illumination with a PC microscope camera (Veho MS-004 Discovery Deluxe USB Microscope). The ventral hindwing was used as its melanization is known to influence heat gain during lateral basking (Heinrich, 1996), whereas dorsal forewing melanization influences the heat gain during dorsal basking (Kingsolver, 1987). To score wing area, we used the 'lasso' function in Adobe Photoshop CS6. Wing melanization was defined as the percentage of black wing area (Figure S2). We used a threshold approach with a fixed value of 128 (on a scale of 0 to 255), turning each pixel on the butterfly wing into either black or white (Adobe Photoshop CS6). We additionally calculated forewing-to-hindwing ratio (forewing area divided by hindwing area), wing loading (total body mass divided by forewing area) and wing aspect ratio to examine wing shape ($4 \times \text{forewing length}^2$ divided by forewing area; Berwaerts et al., 2002).

2.4 | Physiological parameters

We measured the following parameters related to oxidative stress: (a) potential metabolic activity (PMA), (b) two markers of antioxidant defences: total haemolymph antioxidant capacity (OXY) and glutathione (a nonenzymatic antioxidant; GSH) and (c) three markers of oxidative damage: hydroperoxide concentration (a reactive oxygen metabolite; ROM), malondialdehyde concentration (a marker of lipid peroxidation; MDA) and DNA damage (8OHdG). To keep the size of the experiments manageable, PMA, OXY and ROM were only measured in males. Frozen abdomen was cut into two similar-sized parts and weighed (± 0.1 mg). The first half was used to measure PMA, whereas the second half was used to measure OXY and ROM. For measuring GSH, MDA and 8OHdG, we used thoraces and abdomen from other males and females. For details on measuring physiological parameters, see Supplementary Material S3.

3 | Statistical analyses

All data on developmental and morphological traits were analysed using nested ANOVAs with country (Italy, Germany and Sweden), sex, temperature and all resulting interactions as fixed factors, and replicate population as well as family (i.e. the offspring of an individual female) as random effects. Population was nested within country and family within country and population. The physiological

parameters PMA, OXY and ROM were analysed with similar ANOVAs, but without the factors sex (as only males were used here) and family (owing to small sample sizes per family). Likewise, MDA, GSH and DNA damage were analysed using similar ANOVAs, including sex but once again excluding family effects because of small sample sizes per mother. Pupal time was ln-transformed, PMA, ROM and OXY were log-transformed, and DNA damage as well as GSH was 1/x-transformed prior to analyses to meet ANOVA requirements. Models were constructed by stepwise backward removal of nonsignificant interactions (Table S4). Owing to the high number of statistical tests, we corrected all *p*-values for table-wide false discovery rates using the Bonferroni–Holm method (Table 1). To reduce the complexity of the data set, we additionally performed PCAs. Resulting PC scores were analysed with nested ANOVAs as indicated above (Table S5). Throughout the text, means are given as ± 1 SE. All statistical tests were performed with Statistica 12.0 (Tulsa, StatSoft, OK).

4 | RESULTS

4.1 | Developmental traits, morphology and heat knock-down time

After correcting for multiple testing, country significantly affected wing melanization only (Table 1; for details see Table S4).

TABLE 1 Results of nested ANOVAs for the effects of country (fixed factor), population (random, nested within country), family (random, nested within country and population), sex and temperature (both fixed) on various traits in *Pieris napi* from a latitudinal gradient

	Country	Popul	Family	Sex	Temp	Country*Temp
Larval time			***	***	***	***
Pupal time			***		***	
Pupal mass		***	***	***	***	***
Larval growth rate		***		***	***	***
Adult mass			***	***	***	
Thorax mass			***	***	***	
Abdomen mass			***	***	***	***
Thorax/Abdomen				***	***	***
Forewing area		***	***	***	***	
FW melanization	***		***	***	***	***
Hindwing area		***	***	***	***	
HW melanization	***		***	***	***	
FW-HW ratio		***	***	***		
Wing loading			***	***	***	
Wing aspect ratio				***		
Heat tolerance			***	***	***	
PMA					***	

Note: Indicated are only effects being significant after Bonferroni–Holm correction (***). Country-by-sex, sex-by-temperature and three-way interactions were nonsignificant throughout. The following traits were not significantly affected by any factor: OXY (total antioxidant capacity), ROM (reactive oxygen metabolites), malondialdehyde, DNA damage and glutathione. For details on statistical tests, see Table S4.

Abbreviations: FW, forewing; HW, hindwing; PMA, potential metabolic activity.

Forewing and hindwing melanization were higher in Swedish followed by German and finally Italian butterflies ($S > G > I$; Tukey HSD after ANOVA; Figure 2a). Additionally, the PCA revealed that Italian individuals were significantly heavier/larger than German and finally Swedish ones (PC1: $I > G > S$; $F_{2,6} = 13.5$, $p = .0060$; Figure 2b). PC1 was strongly correlated with all measures of body size (all r -values $> .73$; Table S5). Despite being larger, Italian larvae had the shortest development time, realized by higher growth rates compared with German or Swedish larvae (PC2: $I > G = S$; $F_{2,6} = 43.3$, $p = .0003$; Figure 2c). PC2 was positively related to larval time ($r = .713$) but negatively to larval growth rate ($r = -.749$).

Regarding potentially flight-related traits, wing loading (PC2; see above; $r = .729$) and wing aspect ratio (PC5; $F_{2,6} = 8.4$, $p = .0182$; $r = .473$) were higher in German and Swedish than in Italian individuals ($G = S > I$; Figure 2d), whereas thorax-abdomen ratio (PC3, $p = .0869$) did not attain significance. PC6, being strongly related to heat knock-down time ($r = .794$), also differed significantly among countries ($I > G = S$; $F_{2,6} = 5.9$, $p = .0375$; Figure 2e). For several traits, significant differences among populations and families were detected (Table 1).

Significant temperature and sex differences were found for all traits investigated except for pupal time (no sex difference)

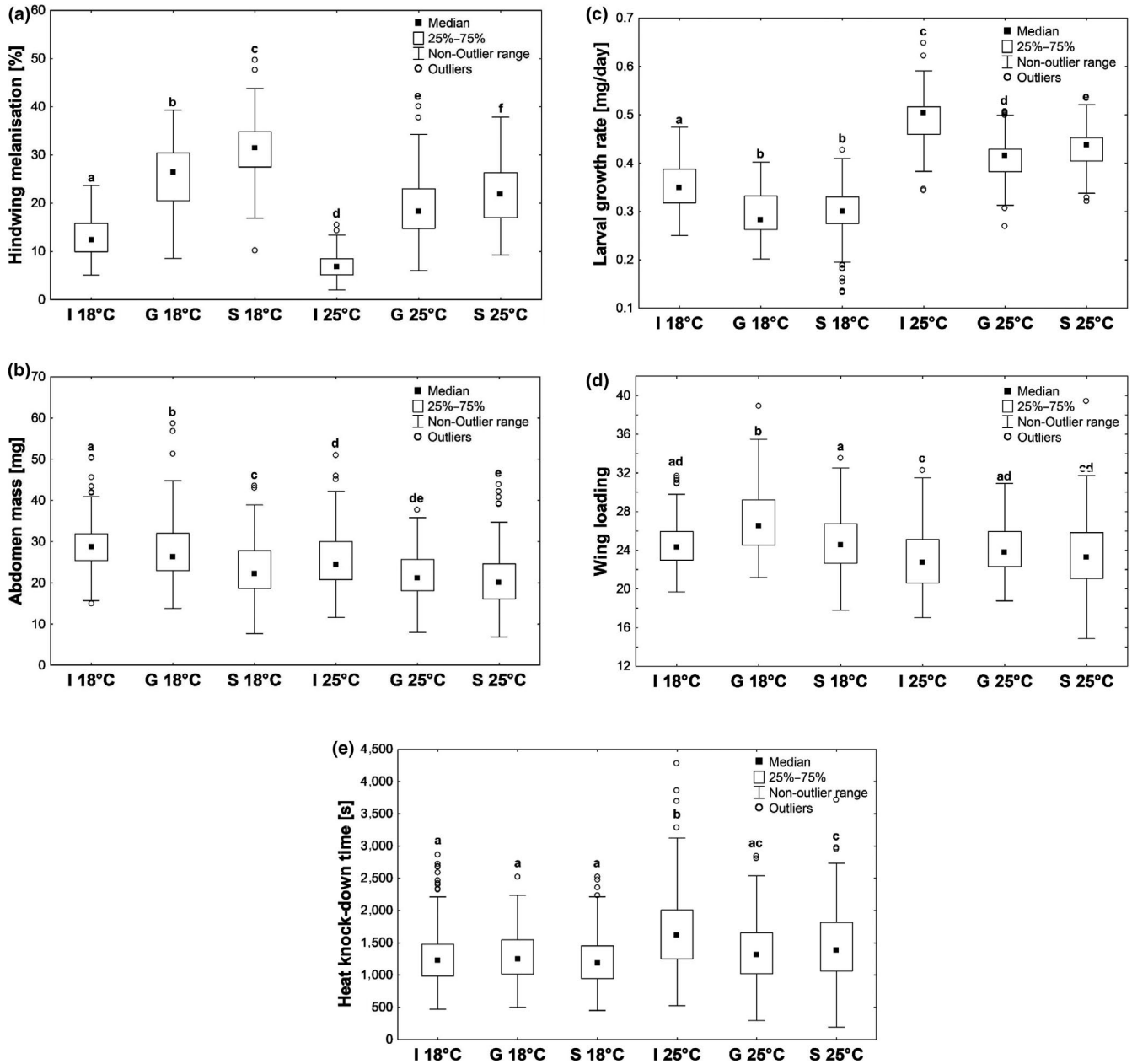


FIGURE 2 Hindwing melanization (a), abdomen mass (b), larval growth rate (c), wing loading (d) and heat knock-down time (e) of *Pieris napi* in relation to latitude (Italy, I; Germany, G; and Sweden, S) and rearing temperature (18°C and 25°C). Different letters above boxes indicate significant differences between groups. All data are pooled across sexes. Group samples sizes range between 90 and 333 individuals for all traits

and forewing-to-hindwing and wing aspect ratio (no temperature difference) (Table 1). Rearing individuals at the higher compared with the lower temperature resulted in shorter larval and pupal development times; higher larval growth rate, thorax–abdomen ratio and heat stress resistance; lower pupal, adult, thorax and abdomen masses, forewing and hindwing melanization and wing loading; and smaller hindwings and forewings (Figure 2; Figure S6). Females had significantly longer larval development times; a lower larval growth rate, pupal and thorax mass, thorax–abdomen ratio, hindwing melanization and wing aspect ratio; smaller forewings and hindwings; and higher adult and abdomen masses, forewing melanization, forewing–hindwing ratio, wing loading and heat stress resistance than males.

The above-mentioned general patterns were partly modulated by interactive effects (Table 1). Country-by-temperature interactions were significant for larval time, pupal mass, larval growth rate, abdomen mass, thorax–abdomen ratio and forewing melanization. In addition, these interactions were often caused by variation in absolute values, while relative changes were similar. For instance, the higher developmental temperature reduced larval time by 4.5 (Italy), 5.1 (Germany) and 5.5 days (Sweden), causing the significant interaction. In relative terms, this is equivalent to a reduction in 30.0, 29.3 and 32.3%. We therefore mention below only those interactions, which reflect differences in relative changes > 5% among countries for a specific trait (see also Figure S6). The higher temperature reduced abdomen mass more strongly in German (–21.8%) than in Italian (–11.6%) or Swedish (–8.8%) individuals (Figure 2b). Moreover, thorax–abdomen ratio increased by 11.0% in German individuals at the higher temperature, whereas it remained virtually unaffected in Italian (–0.4%) or Swedish (–2.7%) individuals. Regarding heat knock-down time, plasticity was highest in Italian animals, showing an increase by 32.1% at the higher temperature compared with 17.9% in Swedish and 3.0% in German individuals (Figure 2f). Note that the latter interaction was not significant after Bonferroni–Holm correction, but well in the PCA (PC6: $F_{2,717} = 11.2$, $p < .0001$). Sex-by-temperature, country-by-sex and three-way interactions were nonsignificant throughout.

4.2 | Physiological traits

Regarding temperature effects, PMA was significantly higher at 18°C ($0.646 \pm 0.017 \mu\text{l O}_2/\text{mg protein/hr}$) than at 25°C ($0.547 \pm 0.019 \mu\text{l O}_2/\text{mg protein/hr}$; Table 1). All other effects were nonsignificant.

5 | DISCUSSION

5.1 | Latitudinal variation

Wing melanization, development time, wing loading and wing aspect ratio increased, whereas body size, larval growth rate and

heat tolerance decreased with increasing latitude. The patterns found for wing melanization, body size and wing loading are in agreement with data based on field-caught individuals from the same cline (Günter et al., 2019). Thus, variation in these traits seems to be buffered across environments. For wing aspect ratio, though, a cline opposite to the one reported earlier was found. This may indicate a large environmental impact on wing aspect ratio. However, overall there was once again no evidence for strong selection on flight-related morphology across latitudes (Günter et al., 2019). For instance, there was no variation in thorax–abdomen ratio and contradictory evidence for wing aspect ratio versus wing loading. Although the increase in wing aspect ratio with latitude is associated with better manoeuvrability in insects, a higher wing loading is clearly detrimental for flight (Berwaerts et al., 2002; Hassall, 2015).

The finding that individuals from warmer rather than cooler environments were larger challenges both the Bergmann's and the temperature-size rule (Angilletta & Dunham, 2003; Chown & Gaston, 1999). As converse Bergmann size clines have been repeatedly reported (Blanckenhorn & Demont, 2004; Blanckenhorn et al., 2006), we suggest that body size is not directly under thermal selection but perhaps more closely associated with season length. Warmer climates are more beneficial for the development of *Pieris* species than cooler ones (Angilletta & Dunham, 2003), probably resulting in the pattern observed. Additionally, German individuals appeared to be more plastic than Italian or Swedish ones with regard to abdomen mass. However, this was not found in other body size-related traits, such that no general conclusions seem possible. Despite being larger, Italian larvae exhibited the shortest larval development time, realized by a higher growth rate compared with German or Swedish ones. Again, this finding challenges our a priori prediction (counter-gradient variation). Thus, populations from warm climates seem to be selected for rapid growth, enabling simultaneously short development time and large size. Perhaps, rapid growth enables squeezing in more generations per year, resulting in compound interest benefits (Fischer & Fiedler, 2002).

The increase in wing melanization towards cooler environments was expected as a darker coloration increases heat absorption, in turn facilitating higher levels of activity (Ellers & Boggs, 2004; Stoehr & Goux, 2008). Therefore, darker individuals will likely more readily reach mating partners, nectar and host plants or escape predators under thermally challenging conditions (Berger, Walters, & Gotthard, 2008). In contrast, strong melanization may cause overheating in warm environments (Kingsolver & Watt, 1983; Van Dyck, Matthysen, & Dhondt, 1997). Thus, clinal variation in melanization results from both, selection to increase it in cooler but to decrease it in warmer environments.

Italian individuals tended to be more heat stress-resistant than German or Swedish ones and also showed higher levels of plasticity in thermal tolerance. Both were expected based on a higher risk of exposure to detrimentally high temperatures in warmer climates in southern Europe. Thus, cold-adapted populations may suffer from future anthropogenic climate change due to both a lower baseline

heat tolerance and low levels of plasticity therein (cf. Karl, Janowitz, & Fischer, 2008). The higher heat resistance of Italian butterflies might be related to higher antioxidant defences, as suggested by a trend towards a higher GSH concentration than in German butterflies. This can prevent oxidative damage caused by reactive oxygen species (Hermes-Lima, 2004; Pamplona & Costantini, 2011). However, the parallel increase in GSH in Swedish animals seems surprising. We speculate that stress imposed by cold temperatures may yield similar effects and therefore adaptations as those imposed by high temperatures (Grim, Simonik, Semones, Kuhn, & Crockett, 2013; Lalouette, Williams, Hervant, Sinclair, & Renault, 2011).

5.2 | Effects of developmental temperature

Higher temperatures generally speed up biochemical processes and metabolic rates, resulting in higher growth rates and thus shorter development times as found here (Angilletta, 2009; Blanckenhorn, 1997; Karl et al., 2008; Kingsolver & Woods, 1997). Likewise, a reduction in body size respectively mass at the higher temperature was expected according to the temperature-size rule, a near-universal 'law' in ectotherms (Angilletta & Dunham, 2003; Atkinson, 1994). The temperature-size rule in turn results from thermal effects on behavioural and physiological mechanisms, namely a reduced food intake and a lower efficiency in converting ingested food into body matter at higher temperatures (Karl & Fischer, 2008).

The reduced wing loading at the higher temperature likely increases flight performance (Berwaerts et al., 2002; Hassall, 2015). Why this should be the case is not obvious, as physical limitations on flight are less pronounced at higher temperatures. Thus, the pattern does not seem to imply adaptive plasticity, but is a consequence of an apparently stronger effect of higher temperatures on body mass than body (wing) size. At the higher temperature, thorax-abdomen ratio was overall higher, indicating that especially abdomen mass and therefore storage reserves were reduced. This likely reflects higher metabolic losses at the higher temperature (Karl & Fischer, 2008; Kingsolver & Woods, 1997). Wing aspect ratio was lower at the higher temperature, which may reduce flying speed and manoeuvrability but increase performance during long-distance flights (Hassall, 2015). The increased wing melanization at lower temperatures likely reflects adaptive plasticity (Karl, Hoffmann, & Fischer, 2010; Peñuelas et al., 2017). Likewise, reduced melanization at higher temperatures may be adaptive for preventing overheating (Kingsolver & Watt, 1983; Van Dyck et al., 1997; Watt, 1968). Individuals reared at the higher temperature showed an increased heat tolerance, reflecting a well-known pattern of adaptive phenotypic plasticity (Fischer & Karl, 2010; Karl et al., 2008).

The decrease in potential metabolic activity at the higher temperature is counterintuitive, though similar patterns have been documented before (e.g. Žagar, Holmstrup, Simčič, Debeljak, & Slotsbo, 2018). The mitochondrial respiratory chain is a major source of reactive oxygen species (ROS), which can cause cellular damage (Hermes-Lima, 2004). The reversible inactivation of enzymes, which

limits enzymatic processes outside the usual temperature range (Simčič, Pajk, Jaklič, Brancelj, & Vrezec, 2014), may additionally decrease PMA. Such a reduction in enzyme activity is adaptive by reducing the risk of cellular damage due to ROS at higher temperatures and may therefore be an 'upstream' antioxidant response to avoid oxidative stress. These antioxidant responses appear efficient, as none of the considered markers of oxidative damage was significantly affected by thermal conditions.

5.3 | Sexual differences

The sexual differences found largely followed expected patterns. Males showed shorter development times, facilitated by higher growth rates, than females, which is typically explained by selection for earlier male emergence to maximize mating opportunities (Fagerström & Wiklund, 1982; Wiklund & Fagerström, 1977). Females had a lower pupal and thorax mass as well as smaller forewings and hindwings than males, reflecting the strong selection on large male body size in *P. napi*, enabling large nuptial gifts (Bergström & Wiklund, 2002). Nevertheless, adult mass was actually higher in females, owing to a much higher mass loss upon metamorphosis in males, which has been described as a cost of the males' rapid development (Fischer, Zeilstra, Hetz, & Fiedler, 2004). Sexual differences in the relative investment into locomotion versus reproduction explain the females' lower thorax-abdomen ratio but higher wing loading (Berwaerts et al., 2002; Karl et al., 2008). With regard to wing coloration, the females' higher degree of forewing melanization results from a second black spot which is missing in males (Tolman & Lewington, 2008). However, hindwing melanization, being relevant to heat absorption (Wasserthal, 1975), was more pronounced in males. This may facilitate flight ability and thus mate location under thermally challenging conditions (Åhman & Karlsson, 2009; Wiklund & Forsberg, 1991).

6 | CONCLUSIONS

Our common garden experiment revealed clinal patterns in wing melanization, body size and wing loading in *P. napi*, which are in strong agreement with field-based data (Günter et al., 2019). Likewise, laboratory and field data concur in that there is no clear evidence for selection on flight-related morphology across latitudes (Günter et al., 2019). Our present data further indicate shorter development times, enabled by higher growth rates, and increased heat tolerance at lower latitudes. These findings suggest that, in *P. napi*, warmer climates (1) are more beneficial for development, (2) select for rapid growth to enable additional generations per year, (3) select for increased heat tolerance (and potentially antioxidant capacities) and (4) relax selection for thermal melanization. Although (3) and (4) likely reflect thermal adaptation, variation in growth trajectories and body size seems to be more closely related to season length. We thus found clear

evidence for genetic adaptation to local conditions. One caveat here is that our study did not control for cross-generational parental effects, but at least all offspring was reared under controlled conditions. Although, in general, parental effects may have important implications (Huestis & Marshall, 2006; Mousseau & Dingle, 1991), their influence is often rather weak as compared with those of the developmental environment (Robinson & Partridge, 2001), in particular in butterflies (Steigenga & Fischer, 2007). Note also that most patterns documented here are in agreement with predictions based on theory and the results of other studies (growth trajectories, Van Doorslaer & Stoks, 2005; heat tolerance, Fischer & Karl, 2010; and wing melanization, Watt, 1968; Van Dyck et al., 1997). In general, plastic responses to temperature were more pronounced than differences among origins, stressing the importance of phenotypic plasticity for dealing with environmental variation. Still, several patterns prevailed under laboratory and field conditions. Some plastic responses were indicative of adaptive phenotypic plasticity, most notably in wing melanization and heat tolerance. Our study highlights that temperature is an important selective agent along latitudinal clines, though the importance of other forces such as season length should not be neglected.

ACKNOWLEDGMENTS

We thank Kristin Franke and Christin Park for help with processing butterflies. This research was funded by the DFG Research Training Group RESPONSE (DFG GRK 2010).

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

K.F. and M.B. conceived the study, K.F., F.G. and M.B. designed the research. F.G., I.W. and K.F.F. performed the experiments. N.T., A.Ž. and T.S. performed the biochemical assays. K.F. and F.G. analysed the data, and K.F., F.G. and M.B. wrote the manuscript.

ORCID

Franziska Günter  <https://orcid.org/0000-0003-4446-6796>

Michaël Beaulieu  <https://orcid.org/0000-0002-9948-269X>

Klaus Fischer  <https://orcid.org/0000-0002-2871-246X>

REFERENCES

- Åhman, M., & Karlsson, B. (2009). Flight endurance in relation to adult age in the green-veined white butterfly *Pieris napi*. *Ecological Entomology*, 34, 783–787. <https://doi.org/10.1111/j.1365-2311.2009.01126.x>
- Angilletta, M. J. J. (2009). *Thermal adaptation: A theoretical and empirical synthesis*, 1st edn. New York, NY: Oxford University Press.
- Angilletta, M. J. J., & Dunham, A. E. (2003). The temperature-size rule in ectotherms: Simple evolutionary explanations may not be general. *The American Society of Naturalists*, 162(3), 332–342. <https://doi.org/10.1086/377187>
- Arendt, J. D. (1997). Adaptive intrinsic growth rates: An integration across taxa. *The Quarterly Review of Biology*, 72(2), 149–177. <https://doi.org/10.1086/419764>
- Arnett, A. E., & Gotelli, N. J. (1999a). Bergmann's rule in the ant lion *Myrmeleon immaculatus* DeGeer (Neuroptera: Myrmeleontidae): Geographic variation in body size and heterozygosity. *Journal of Biogeography*, 26, 275–283. <https://doi.org/10.1046/j.1365-2699.1999.00271.x>
- Arnett, A. E., & Gotelli, N. J. (1999b). Geographic variation in life-history traits of the ant lion, *Myrmeleon immaculatus*: Evolutionary implications of Bergmann's rule. *Evolution*, 53(4), 1180–1188. <https://doi.org/10.2307/2640821>
- Atkinson, D. (1994). Temperature and organism size—a biological law for ectotherms? In M. Begon & A. H. Fitter (Eds.), *Advances in ecological research*, Vol. 25 (pp. 1–58). San Diego, CA: Academic Press Inc. [https://doi.org/10.1016/S0065-2504\(08\)60212-3](https://doi.org/10.1016/S0065-2504(08)60212-3)
- Berger, D., Walters, R., & Gotthard, K. (2008). What limits insect fecundity? Body size- and temperature-dependent egg maturation and oviposition in a butterfly. *Functional Ecology*, 22, 523–529. <https://doi.org/10.1111/j.1365-2435.2008.01392.x>
- Bergström, J., & Wiklund, C. (2002). Effects of size and nuptial gifts on butterfly reproduction: Can females compensate for a smaller size through male-derived nutrients? *Behavioral Ecology and Sociobiology*, 52, 296–302. <https://doi.org/10.1007/s00265-002-0512-0>
- Berwaerts, K., Van Dyck, H., & Aerts, P. (2002). Does flight morphology relate to flight performance? An experimental test with the butterfly *Pararge aegeria*. *Functional Ecology*, 16, 484–491. <https://doi.org/10.1046/j.1365-2435.2002.00650.x>
- Blanckenhorn, W. U. (1997). Altitudinal life history variation in the dung flies *Scathophaga stercoraria* and *Sepsis cynipsea*. *Oecologia*, 109, 342–352. <https://doi.org/10.1007/s004420050092>
- Blanckenhorn, W. U., & Demont, M. (2004). Bergmann and converse Bergmann latitudinal clines in arthropods: Two ends of a continuum? *Integrative and Comparative Biology*, 44, 413–424. <https://doi.org/10.1093/icb/44.6.413>
- Blanckenhorn, W. U., Stillwell, R. C., Young, K. A., Fox, C. W., & Ashton, K. G. (2006). When Rensch meets Bergmann: Does sexual size dimorphism change systematically with latitude? *Evolution*, 60(10), 2004–2011. <https://doi.org/10.1554/06-110.1>
- Bradshaw, W. E., & Holzapfel, C. M. (2008). Genetic response to rapid climate change: It's seasonal timing that matters. *Molecular Ecology*, 17, 157–166. <https://doi.org/10.1111/j.1365-294X.2007.03509.x>
- Chown, S. L., & Gaston, K. J. (1999). Exploring links between physiology and ecology at macro-scales: The role of respiratory metabolism in insects. *Biological Reviews*, 74, 87–120. <https://doi.org/10.1111/j.1469-185X.1999.tb00182.x>
- Chown, S. L., & Klok, J. C. (2003). Altitudinal body size clines: Latitudinal effects associated with changing seasonality. *Ecography*, 26, 445–455. <https://doi.org/10.1034/j.1600-0587.2003.03479.x>
- Crispo, E. (2008). Modifying effects of phenotypic plasticity on interactions among natural selection, adaptation and gene flow. *Journal of Evolutionary Biology*, 21, 1460–1469. <https://doi.org/10.1111/j.1420-9101.2008.01592.x>
- Dahlggaard, J., Hasson, E., & Loeschcke, V. (2001). Behavioral differentiation in oviposition activity in *Drosophila buzzatii* from highland and lowland populations in Argentina: Plasticity or thermal adaptation? *Evolution*, 55(4), 738–747. [https://doi.org/10.1554/0014-3820\(2001\)055\[0738:bdioai\]2.0.co;2](https://doi.org/10.1554/0014-3820(2001)055[0738:bdioai]2.0.co;2)
- Ebert, G., & Rennwald, E. (1993). *Die Schmetterlinge Baden-Württembergs Vol 1: Tagfalter 1*, 1st edn. Stuttgart, Germany: Ulmer.
- Ellers, J., & Boggs, C. L. (2004). Functional ecological implications of intraspecific differences in wing melanization in *Colias* butterflies. *Biological Journal of the Linnean Society*, 82, 79–87. <https://doi.org/10.1111/j.1095-8312.2004.00319.x>
- Fagerström, T., & Wiklund, C. (1982). Why do males emerge before females? Protandry as a mating strategy in male and female butterflies. *Oecologia*, 52, 164–466.

- Fischer, K., & Fiedler, K. (2002). Reaction norms for age and size at maturity in response to temperature: A test of the compound interest hypothesis. *Evolutionary Ecology*, *16*, 333–349. <https://doi.org/10.2307/3545787>
- Fischer, K., & Karl, I. (2010). Exploring plastic and genetic responses to temperature variation using copper butterflies. *Climate Research*, *43*, 315. <https://doi.org/10.3354/cr00892>
- Fischer, K., Zeilstra, I., Hetz, S. K., & Fiedler, K. (2004). Physiological costs of growing fast: Does accelerated growth reduce pay-off in adult fitness? *Evolutionary Ecology*, *18*, 343–353. <https://doi.org/10.1007/s10682-004-2004-3>
- Fox, R., Brereton, T., Asher, J., Botham, M. S., Middlebrook, I., Roy, D. B., & Warren, M. S. (2015). *The state of the UK's butterflies 2015*. Wareham, Dorset: Butterfly Conservation.
- Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M., & Charnov, E. L. (2001). Effects of size and temperature on metabolic rate. *Science*, *293*, 2248–2251. <https://doi.org/10.1126/science.1061967>
- Grim, J. M., Simonik, E. A., Semones, M. C., Kuhn, D. E., & Crockett, E. L. (2013). The glutathione-dependent system of antioxidant defense is not modulated by temperature acclimation in muscle tissues from striped bass, *Morone saxatilis*. *Comparative Biochemistry and Physiology - Part A*, *164*, 383–390. <https://doi.org/10.1016/j.cbpa.2012.11.018>
- Günter, F., Beaulieu, M., Brunetti, M., Lange, L., Schmitz Ornés, A., & Fischer, K. (2019). Latitudinal and altitudinal variation in ecologically important traits in a widespread butterfly. *Biological Journal of the Linnean Society*, *20*, 1–14.
- Hassall, C. (2015). Strong longitudinal variation in wing aspect ratio of a damselfly, *Calopteryx maculata* (Odonata: Zygoptera). *PeerJ*, *3*, e1219. <https://doi.org/10.7717/peerj.1219>
- Heinrich, B. (1996). *The thermal warriors: Strategies of insect survival*. London, UK: Harvard University Press.
- Hermes-Lima, M. (2004). Oxygen in biology and biochemistry: Role of free radicals. In K. B. Storey (Ed.), *Functional metabolism: regulation and adaptation* (pp. 319–368). Hoboken, NJ: Wiley and Sons, Inc. [https://doi.org/10.1016/0160-9327\(93\)90080-m](https://doi.org/10.1016/0160-9327(93)90080-m)
- Hoffmann, A. A., & Rieseberg, L. H. (2008). Revisiting the impact of inversions in evolution: From population genetic markers to drivers of adaptive shifts and speciation? *Annual Review of Ecology, Evolution and Systematics*, *39*, 21–42. <https://doi.org/10.1146/annurev.ecolsys.39.110707.173532>
- Huestis, D. L., & Marshall, J. L. (2006). Interaction between maternal effects and temperature affects diapause occurrence in the cricket *Allonemobius socius*. *Oecologia*, *146*(4), 513–520. <https://doi.org/10.1007/s00442-005-0232-z>
- Ju, R., Wei, H.-P., Wang, F., Zhou, X.-H., & Li, B. (2014). Anaerobic respiration and antioxidant responses of *Corythucha ciliata* (Say) adults to heat-induced oxidative stress under laboratory and field conditions. *Cell Stress and Chaperones*, *19*, 255–262. <https://doi.org/10.1007/s12192-013-0451-x>
- Kapun, M., Fabian, D. K., Goudet, J., & Flatt, T. (2016). Genomic evidence for adaptive inversion clines in *Drosophila melanogaster*. *Molecular Biology and Evolution*, *33*(5), 1317–1336. <https://doi.org/10.1093/molbev/msw016>
- Karl, I., & Fischer, K. (2008). Why get big in the cold? Towards a solution to a life-history puzzle. *Oecologia*, *155*, 215–225. <https://doi.org/10.1007/s00442-007-0902-0>
- Karl, I., Hoffmann, K. H., & Fischer, K. (2010). Cuticular melanisation and immune response in a butterfly: local adaptation and lack of correlation. *Ecological Entomology*, *35*, 523–528.
- Karl, I., Janowitz, S. A., & Fischer, K. (2008). Altitudinal life-history variation and thermal adaptation in the copper butterfly *Lycaena tityrus*. *Oikos*, *117*, 778–788.
- Kawecki, T. J., & Ebert, D. (2004). Conceptual issues in local adaptation. *Ecology Letters*, *7*, 1225–1241. <https://doi.org/10.1111/j.1461-0248.2004.00684.x>
- Kingsolver, J. G. (1987). Evolution and coadaptation of thermoregulatory behavior and wing pigmentation pattern in Pierid butterflies. *Evolution*, *41*(3), 472–490. <https://doi.org/10.2307/2409250>
- Kingsolver, J. G., & Watt, W. B. (1983). Thermoregulatory strategies in *Colias* butterflies: Thermal stress and the limits to adaptation in temporally varying environments. *The American Naturalist*, *121*(1), 32–55. <https://doi.org/10.1086/284038>
- Kingsolver, J. G., & Woods, H. A. (1997). Thermal sensitivity of growth and feeding in *Manduca sexta* caterpillars. *Physiological Zoology*, *70*(6), 631–638. <https://doi.org/10.1086/515872>
- Lalouette, L., Williams, C. M., Hervant, F., Sinclair, B. J., & Renault, D. (2011). Metabolic rate and oxidative stress in insects exposed to low temperature thermal fluctuations. *Comparative Biochemistry and Physiology. Part A*, *158*, 229–234. <https://doi.org/10.1016/j.cbpa.2010.11.007>
- Mousseau, T. A., & Dingle, H. (1991). Maternal effects in insect life histories. *Annual Review of Entomology*, *36*, 511–534.
- Norry, F. M., Bublly, O. A., & Loeschcke, V. (2001). Developmental time, body size and wing loading in *Drosophila buzzatii* from lowland and highland populations in Argentina. *Hereditas*, *135*, 35–40. <https://doi.org/10.1111/j.1601-5223.2001.t01-1-00035.x>
- Pamplona, R., & Costantini, D. (2011). Molecular and structural antioxidant defenses against oxidative stress in animals. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, *301*(4), R843–R863. <https://doi.org/10.1152/ajpregu.00034.2011>
- Parmesan, C. (2006). Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution and Systematics*, *37*, 637–669. <https://doi.org/10.1146/annurev.ecolsys.37.091305.110100>
- Parmesan, C., Ryrholm, N., Stefanescu, C., Hill, J. K., Thomas, C. D., Descimon, H., ... Warren, M. (1999). Poleward shifts in geographical ranges of butterfly species associated with regional warming. *Nature*, *399*, 579–583. <https://doi.org/10.1038/21181>
- Peñuelas, J., Sardans, J., Filella, I., Estiarte, M., Llusà, J., Ogaya, R., ... Terradas, J. (2017). Impacts of global change on Mediterranean forests and their services. *Forests*, *8*, 1–37. <https://doi.org/10.3390/f8120463>
- Reeve, M. W., Fowler, K., & Partridge, L. (2000). Increased body size confers greater fitness at lower experimental temperature in male *Drosophila melanogaster*. *Journal of Evolutionary Biology*, *13*, 836–844. <https://doi.org/10.1046/j.1420-9101.2000.00216.x>
- Robinson, S. J. W., & Partridge, L. (2001). Temperature and clinal variation in larval growth efficiency in *Drosophila melanogaster*. *Journal of Evolutionary Biology*, *14*, 14–21. <https://doi.org/10.1046/j.1420-9101.2001.00259.x>
- Sambucetti, P., Loeschcke, V., & Norry, F. M. (2006). Developmental time and size-related traits in *Drosophila buzzatii* along an altitudinal gradient from Argentina. *Hereditas*, *143*, 77–83. <https://doi.org/10.1111/j.2006.0018-0661.01934.x>
- Simčić, T., Pajk, F., Jaklič, M., Brancelj, A., & Vrezec, A. (2014). The thermal tolerance of crayfish could be estimated from respiratory electron transport system activity. *Journal of Thermal Biology*, *41*, 21–30. <https://doi.org/10.1016/j.jtherbio.2013.06.003>
- Steigenga, M. J., & Fischer, K. (2007). Within- and between-generation effects of temperature on life-history traits in a butterfly. *Journal of Thermal Biology*, *32*, 396–405.
- Stoehr A. M., Goux H.. Seasonal phenotypic plasticity of wing melanisation in the cabbage white butterfly, *Pieris rapae* L. (Lepidoptera: Pieridae). *Ecological Entomology*, 2008; *33*(1), 137-143.
- Tolman, T., & Lewington, R. (2008). *Collins butterfly guide*, 3rd edn. London, UK: HarperCollins Publishers.
- Tumminello, R. A., & Fuller-Espie, S. L. (2013). Heat stress induces ROS production and histone phosphorylation in celomocytes of *Eisenia hortensis*. *Invertebrate Survival Journal*, *10*, 50–57.
- Van Doorslaer, W., & Stoks, R. (2005). Growth rate plasticity to temperature in two damselfly species differing in latitude: Contributions

- of behaviour and physiology. *Oikos*, 111, 599–605. <https://doi.org/10.1111/j.1600-0706.2005.14335.x>
- Van Dyck, H., Matthysen, E., & Dhondt, A. A. (1997). The effect of wing colour on male behavioural strategies in the speckled wood butterfly. *Animal Behaviour*, 53, 39–51. <https://doi.org/10.1006/anbe.1996.0276>
- Wasserthal, L. T. (1975). The rôle of butterfly wings in regulation of body temperature. *Journal of Insect Physiology*, 21, 1921–1930. [https://doi.org/10.1016/0022-1910\(75\)90224-3](https://doi.org/10.1016/0022-1910(75)90224-3)
- Watt, W. B. (1968). Adaptive significance of pigment polymorphism in *Colias* butterflies. I. Variation of melanin pigment in relation to thermoregulation. *Evolution*, 22(3), 437–458. <https://doi.org/10.1111/j.1558-5646.1968.tb03985.x>
- Wiklund, C., & Fagerström, T. (1977). Why do males emerge before females? *Oecologia*, 31, 153–158. <https://doi.org/10.1007/bf00346917>
- Wiklund, C., & Forsberg, J. (1991). Sexual size dimorphism in relation to female polygamy and protandry in butterflies: A comparative study of Swedish Pieridae and Satyridae. *Oikos*, 60(3), 373–381. <https://doi.org/10.2307/3545080>
- Wiklund, C., & Kaitala, A. (1995). Sexual selection for large male size in a polyandrous butterfly: The effect of body size on male versus female reproductive success in *Pieris napi*. *Behavioral Ecology*, 6, 6–13. <https://doi.org/10.1093/beheco/6.1.6>
- Žagar, A., Carretero, M. A., Marguč, D., Simčič, T., & Vrezec, A. (2018). A metabolic syndrome in terrestrial ectotherms with different elevational and distribution patterns. *Ecography*, 41, 1728–1739. <https://doi.org/10.1111/ecog.03411>
- Žagar, A., Holmstrup, M., Simčič, T., Debeljak, B., & Slotsbo, S. (2018). Comparing *Enchytraeus albidus* populations from contrasting climatic environments suggest a link between cold tolerance and metabolic activity. *Comparative Biochemistry and Physiology*, 224, 35–41. <https://doi.org/10.1016/j.cbpa.2018.05.020>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Günter F, Beaulieu M, Freiberg KF, et al. Genotype–environment interactions rule the response of a widespread butterfly to temperature variation. *J Evol Biol*. 2020;33:920–929. <https://doi.org/10.1111/jeb.13623>